

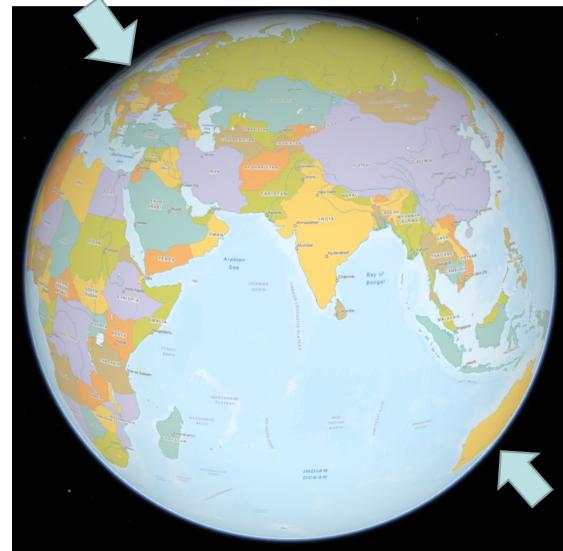


MONASH University

Institute of Pharmaceutical Sciences



UNIVERSITY OF
COPENHAGEN



SAXS on *(food related)* soft matter during digestion

Ben J. Boyd and many others

Department of Pharmacy – University of Copenhagen

Monash Institute of Pharmaceutical Sciences

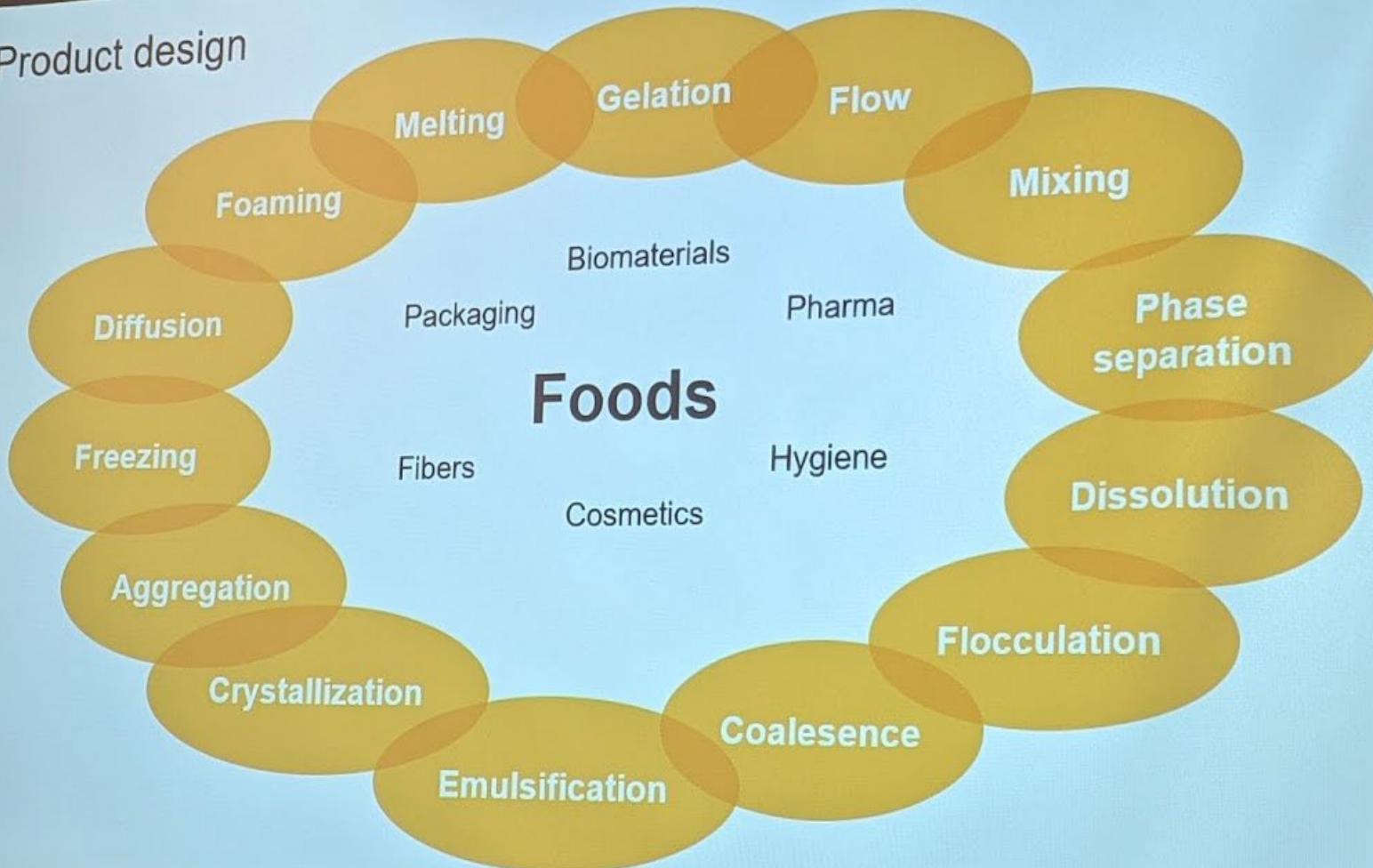
ben.boyd@sund.ku.dk

LINXS Masterclass November 2021

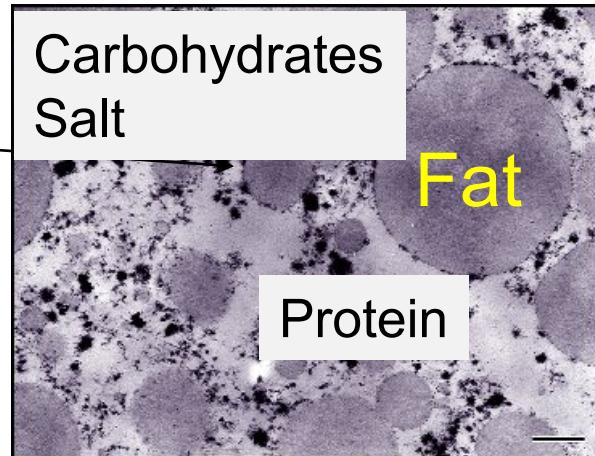
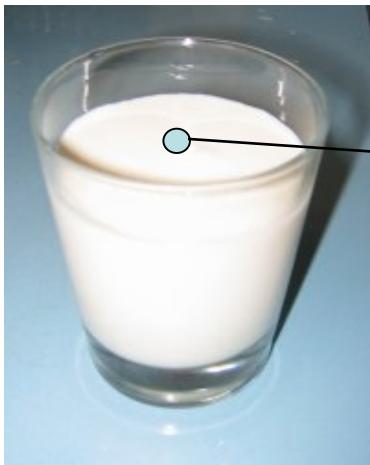


@group_boyd or nonlamellar.com

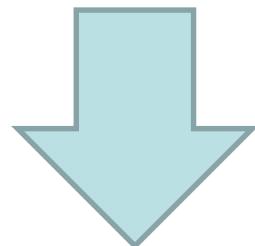
Product design



RI
SE

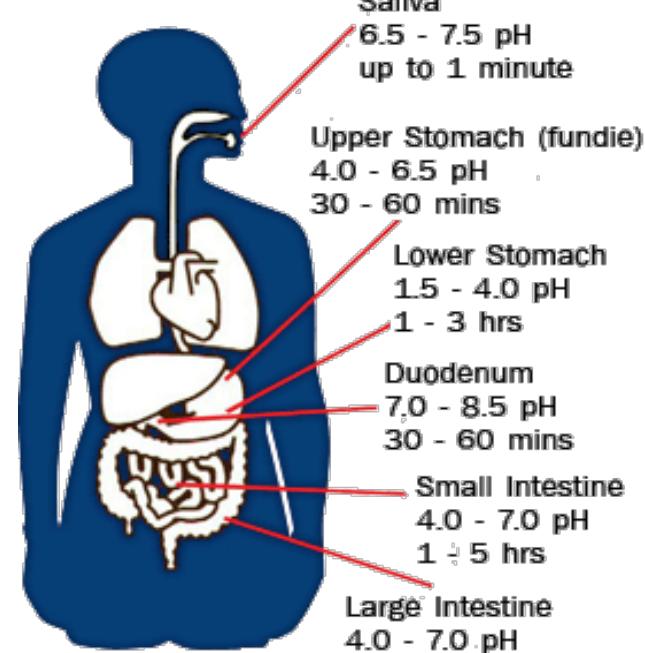


Consider milk as a prototypical food!



Digestion

Temperature
pH
Processing history



This diagram illustrates the average time food spends in each part of the digestive system along with the average pH.

Enzyme

Lipases

Phospholipases

Amylase

Lactase

Proteases

Substrate

Triglycerides – monoglycerides and fatty acids

Phospholipids – lysophospholipids + fatty acids

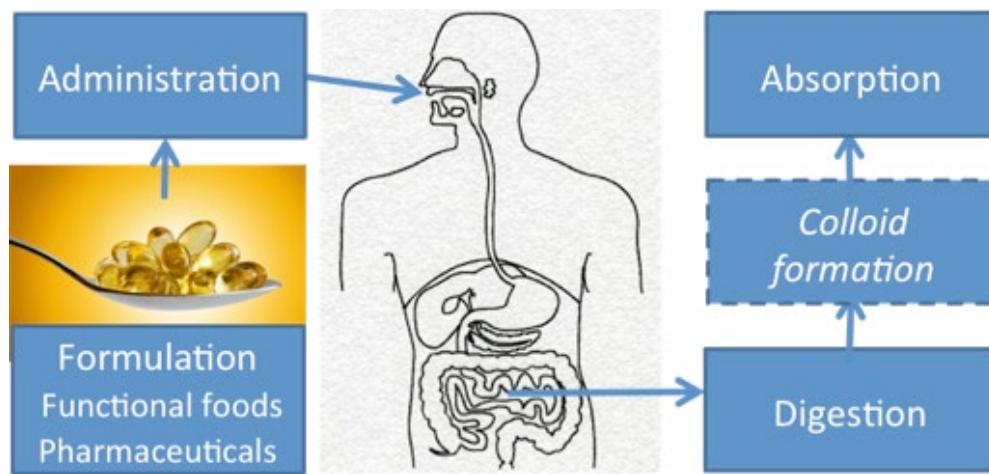
Carbohydrates (starch, alginates) – starch fragments/monosaccharides

Lactose – galactose and glucose

Proteins e.g. casein – peptides and amino acids

Products

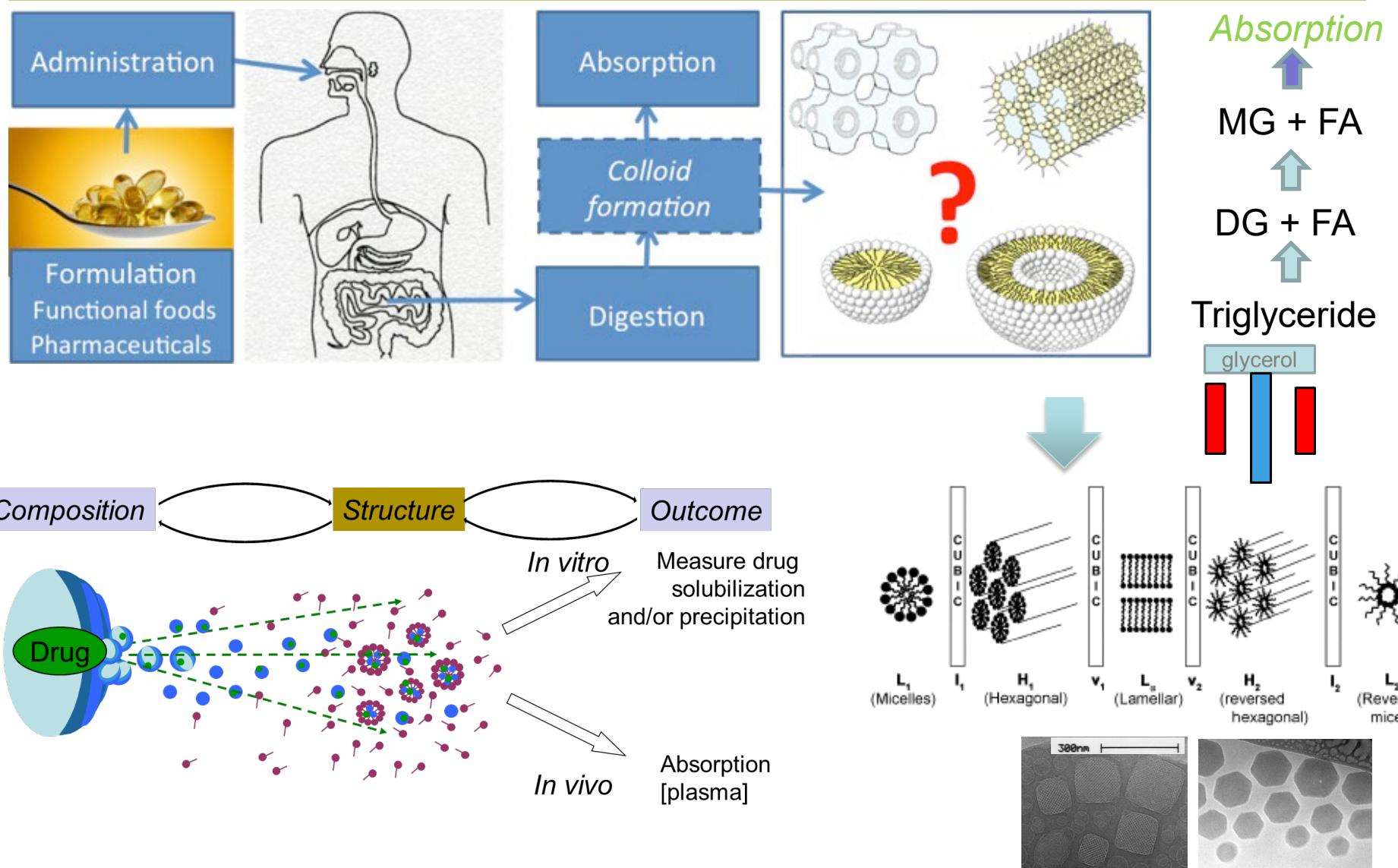
Digestion may lead to changes in structure



Changes in structure due to digestion may lead to:

1. Changes in rate of absorption of nutrients (pharmacokinetics)
2. Changes in extent of absorption of nutrients (bioavailability)
3. Changes in reaction of the body to food (e.g. immunogenicity)
4. Separation of components (demixing, coacervation)
5. Degradation/enhanced stability (unintended release)

Structure in the context of lipids – hierarchical structure



Lipase added to MO cubic phase

Effect of Lipase on Different Lipid Liquid Crystalline Phases Formed by Oleic Acid Based Acylglycerols in Aqueous Systems

100 mM phosphate,

starting pH = 8

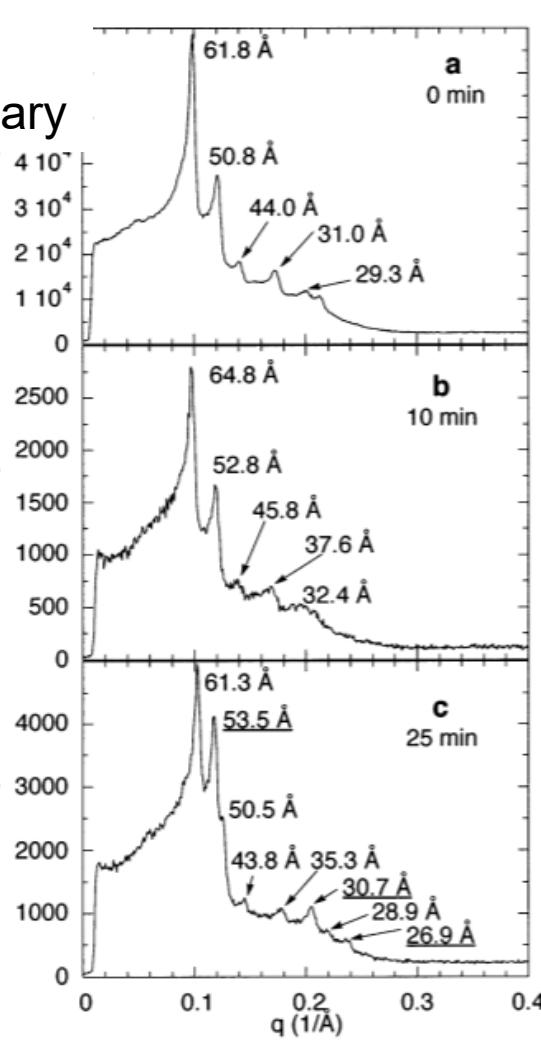
Fixed sample in capillary

V_2

V_2

V_2

Intensity



Johanna Borné, Tommy Nylander,* and Ali Khan

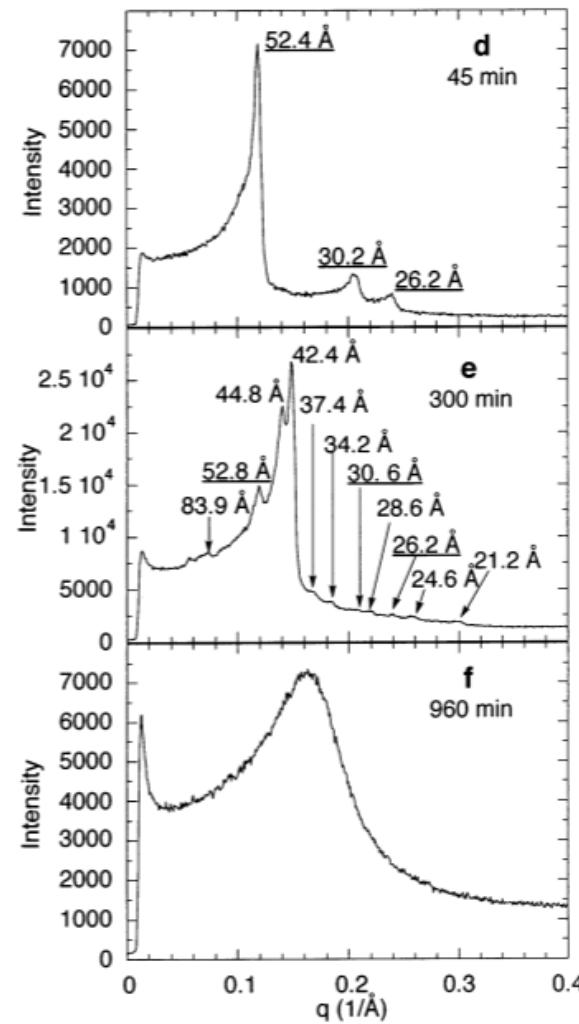
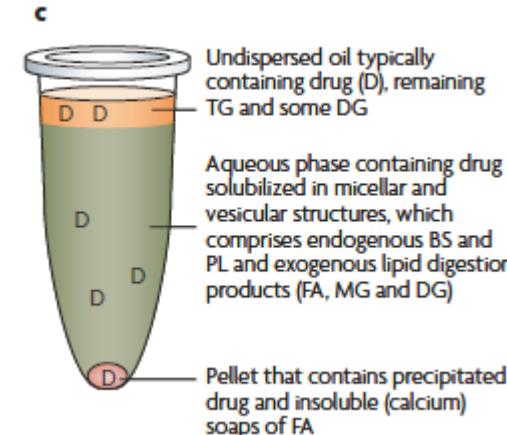
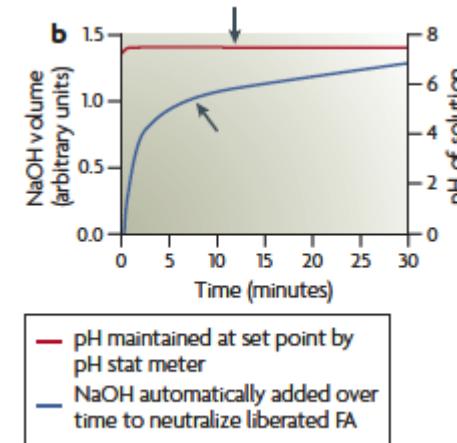
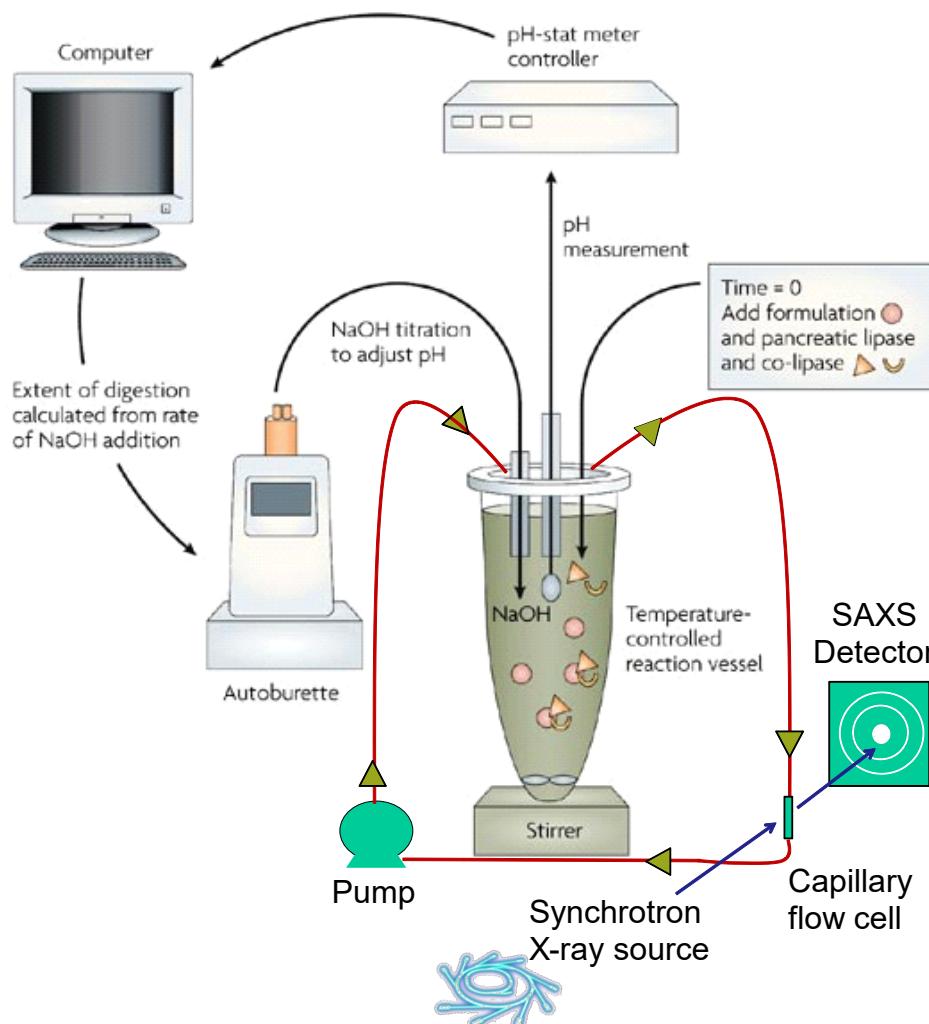


Figure 2. SAXD data recorded after adding 20 μ L of (512 nM) TLL solution to 30 mg of C_D phase (63 wt % MO, 37 wt % $^2\text{H}_2\text{O}$) at 25 $^\circ\text{C}$, giving a final TLL concentration of 205 nM. The intensity is plotted versus the wave vector, $q = 2\pi/d$, where d is the spacing between the lattice planes. The sequence of phase transitions observed is (a) C_D (0 min), (b) C_D (10 min), (c) $C_D + H_{II}$ (25 min), (d) H_{II} (45 min), (e) $C_{mic} + H_{II}$ (300 min), and (f) L_2 (960 min). The d values for the reflections corresponding to the H_{II} phase have been underlined in the figure.

Lipid digestion model with time resolved scattering

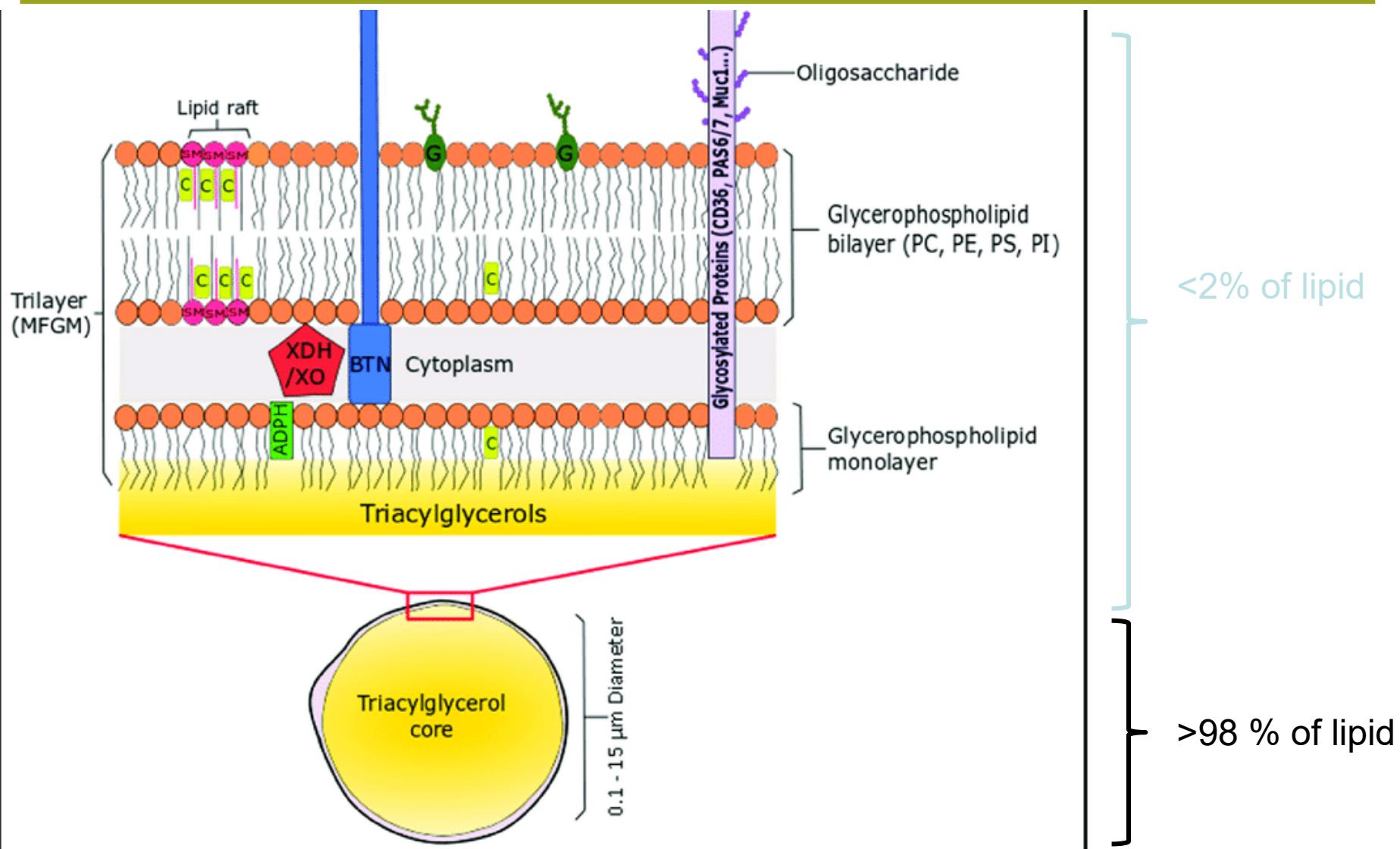


Time-resolved structural information during digestion

Figure from Porter, C.J.H., N.L. Trevaskis, and W.N. Charman, *Lipids and lipid-based formulations: optimizing the oral delivery of lipophilic drugs*. *Nature Reviews Drug Discovery*, 2007. **6**(3): p. 231-248.

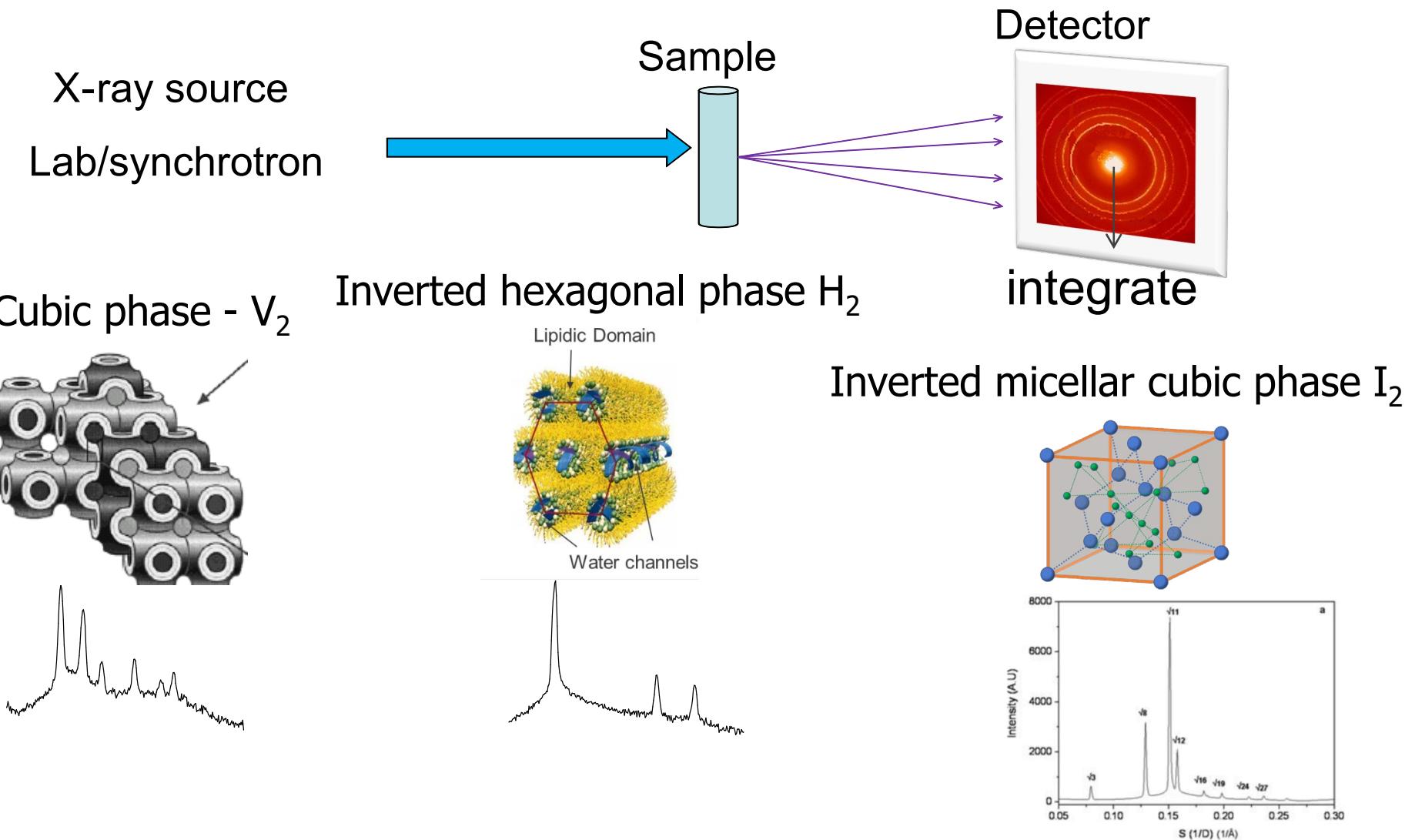


'Structure' of milk fat droplets



- Anticipate triglycerides will dominate structural changes upon digestion?

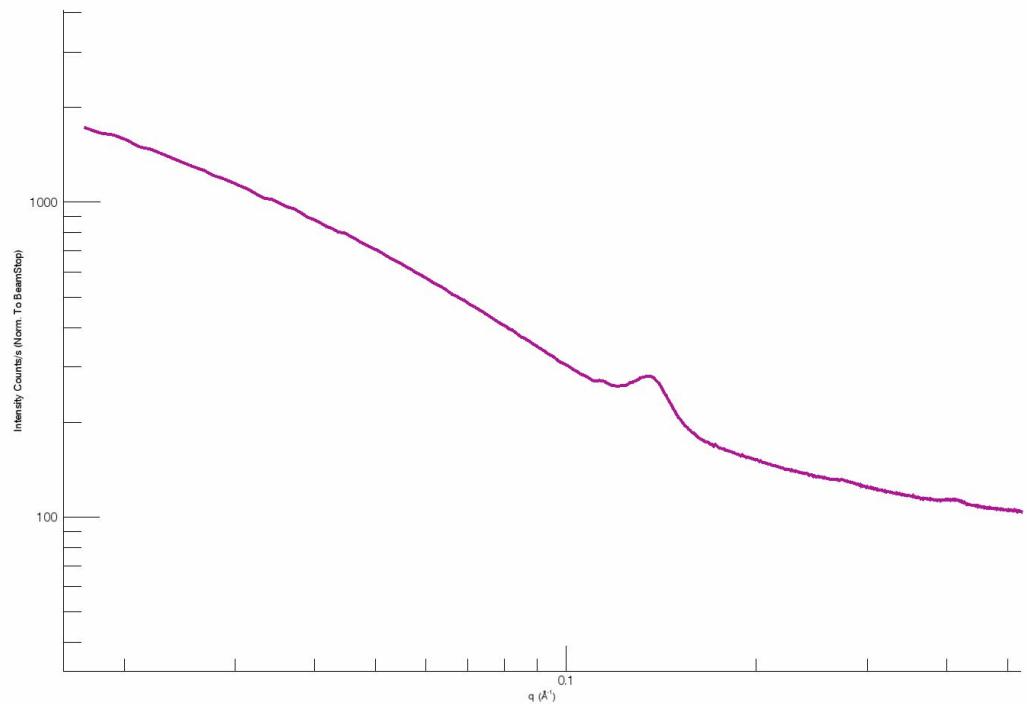
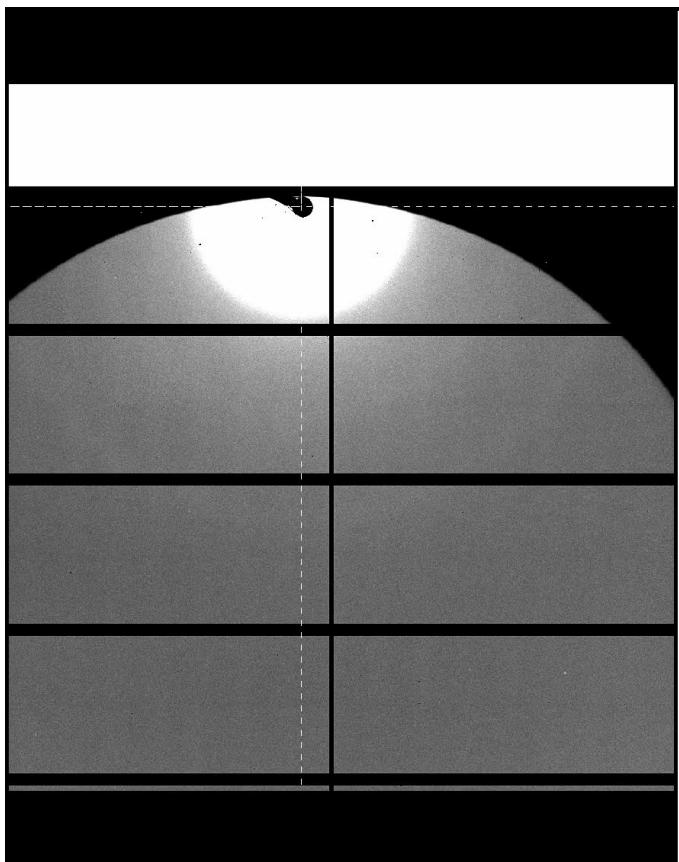
Characterization – small angle X-ray scattering (SAXS)



Scattering is a ‘fingerprint’ for internal phase structure

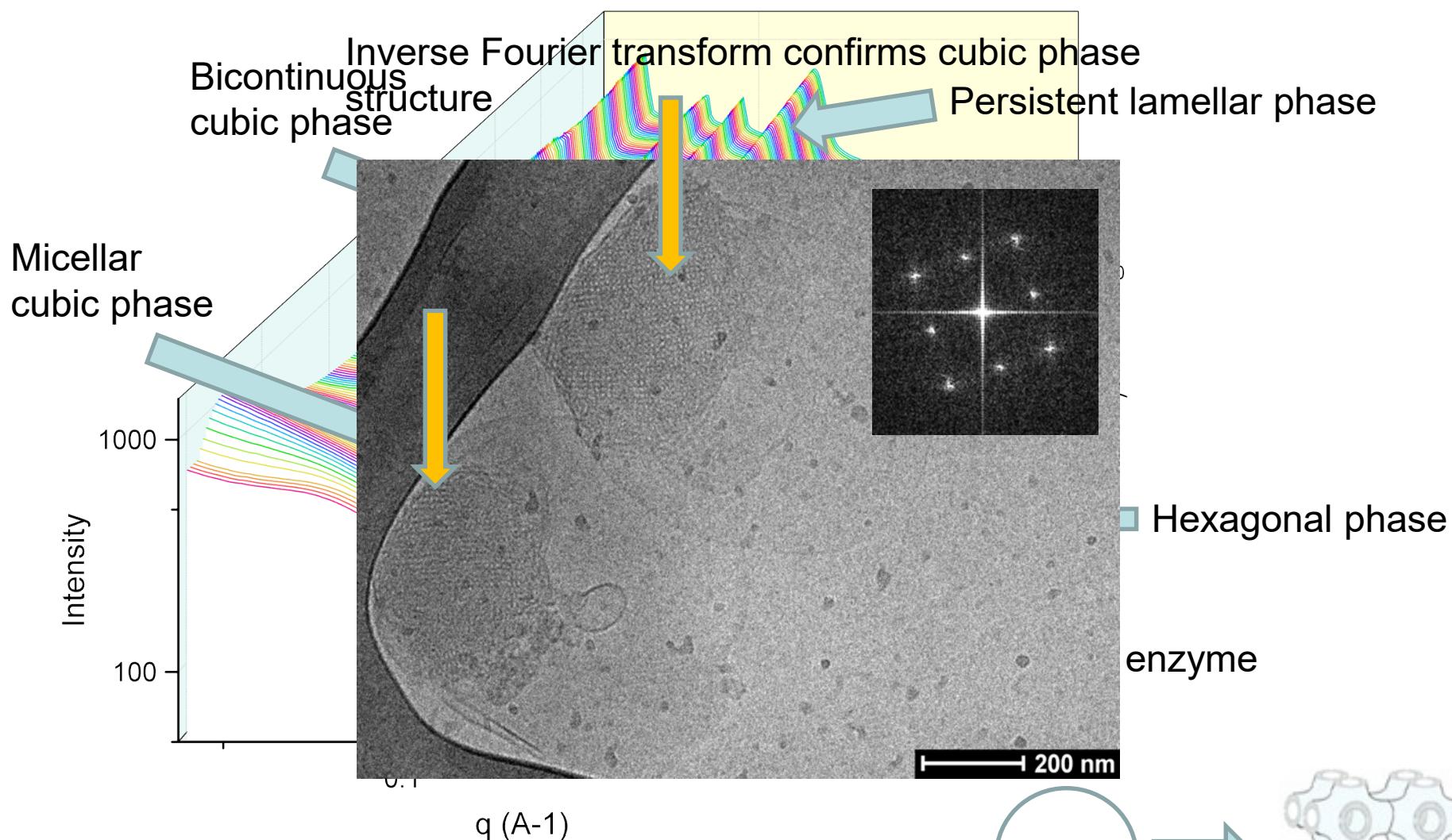
What do we 'see'?

Milk ! – The Movie

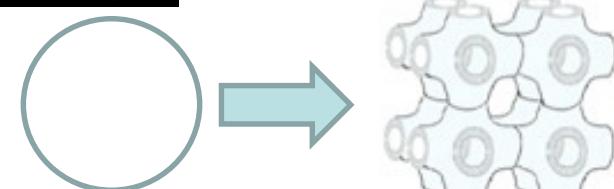


- Complex dynamic transformations in structure

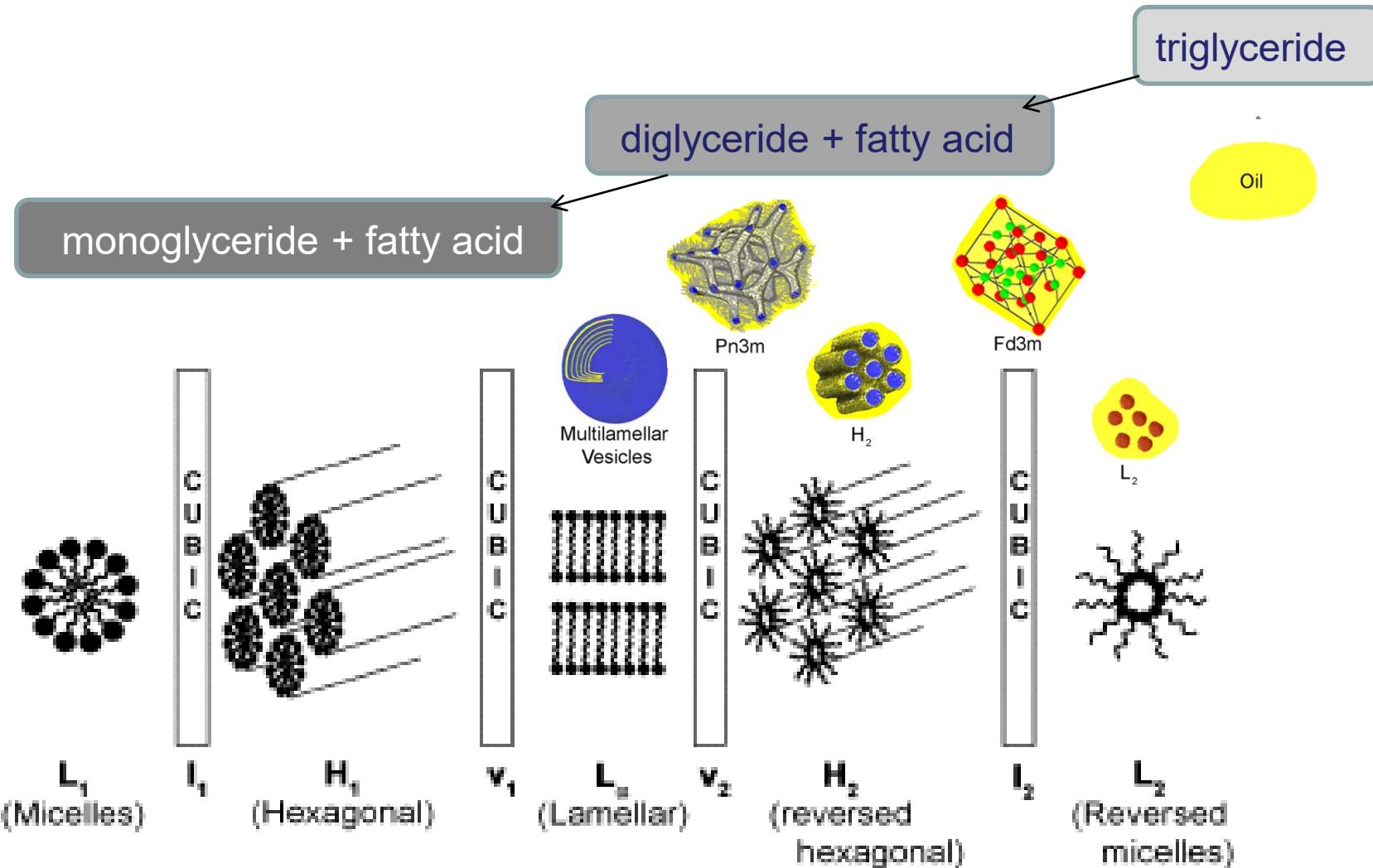
What do we 'see' when we digest milk?



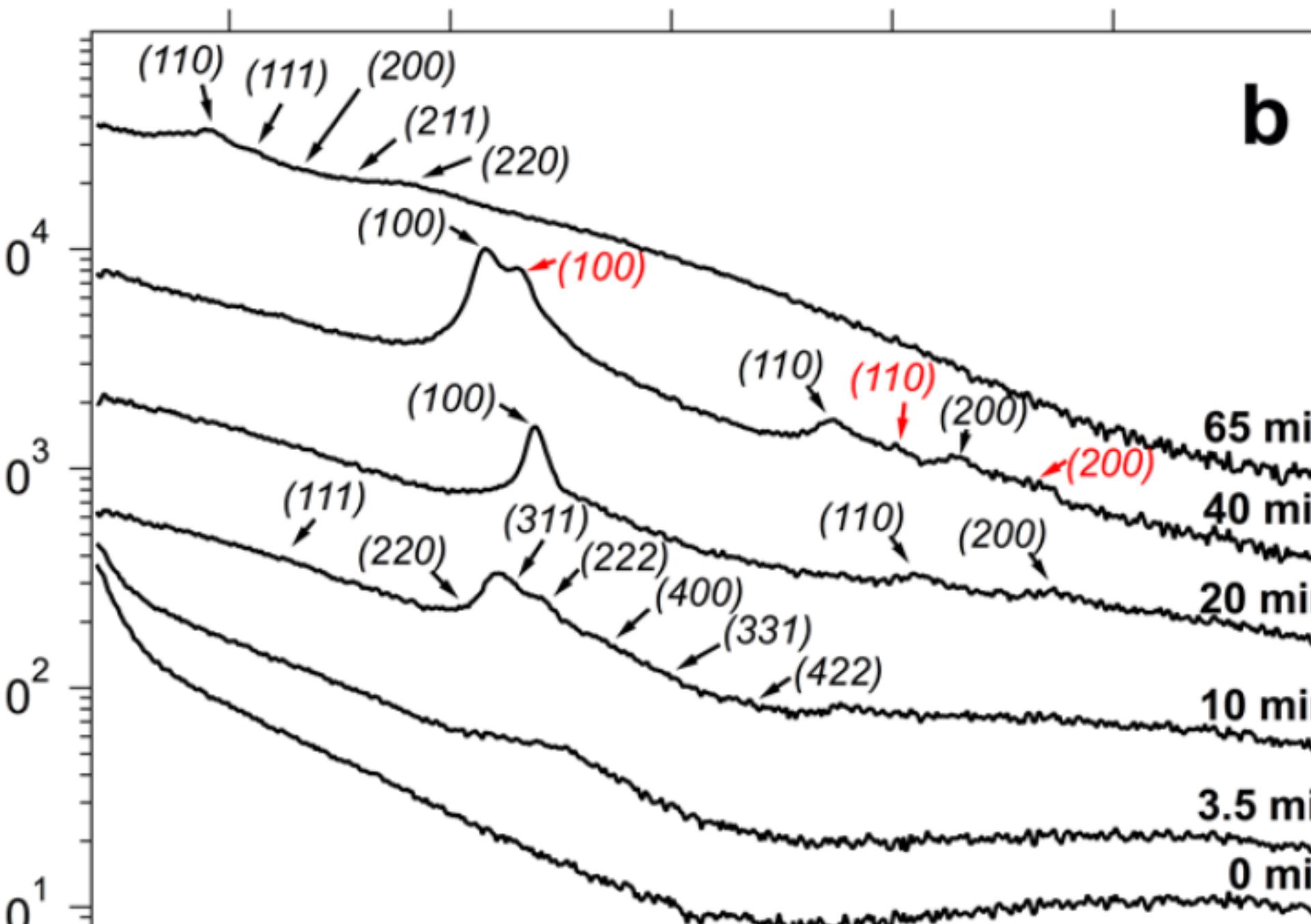
- Structures formed from unstructured emulsion
- Highly dynamic system during digestion
- Droplets transform from liquid to 'cubosomes'



Digestion induces systematic change in lipid packing



“Makes sense” from a physical chemistry/packing parameter perspective

b

Krill oil and astaxanthin (high ω -3 TG) oil

- Krill oil (crude mixture) contains phospholipids, self-stabilising on dispersion
- Astaxanthin oil is purified TG fraction, emulsified in casein or citrem
- Same model as before, SAXS at PSI
- Simplicity removed the structure formation

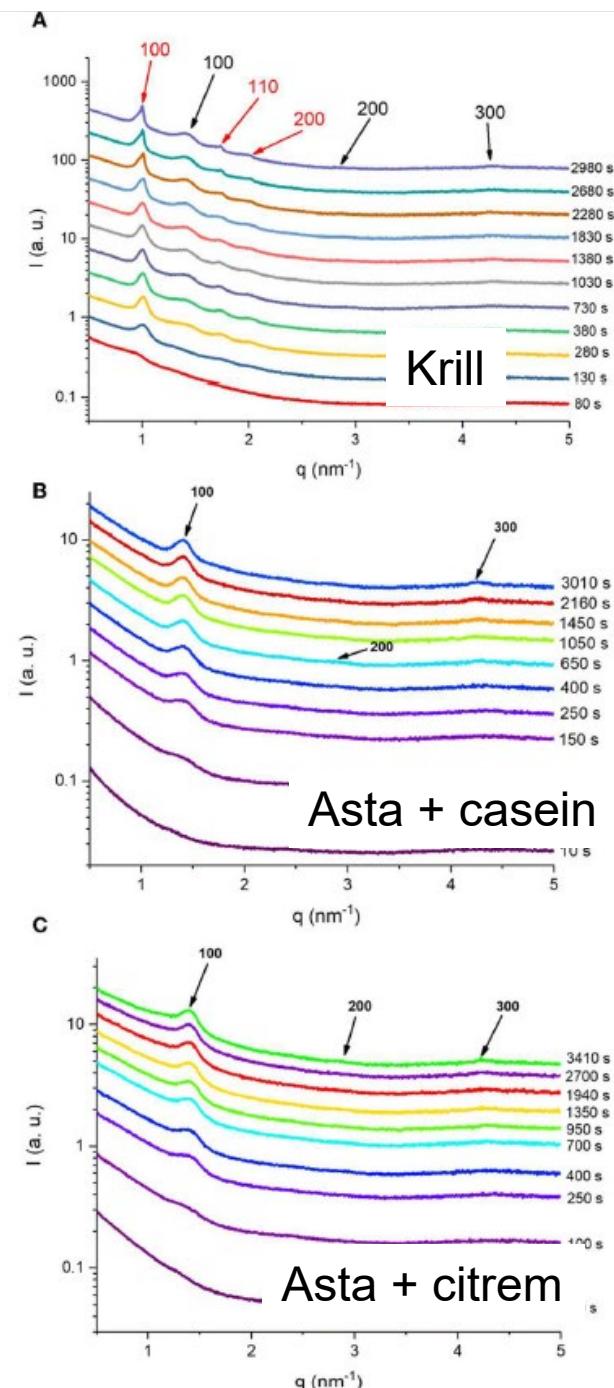


in Bioengineering and Biotechnology

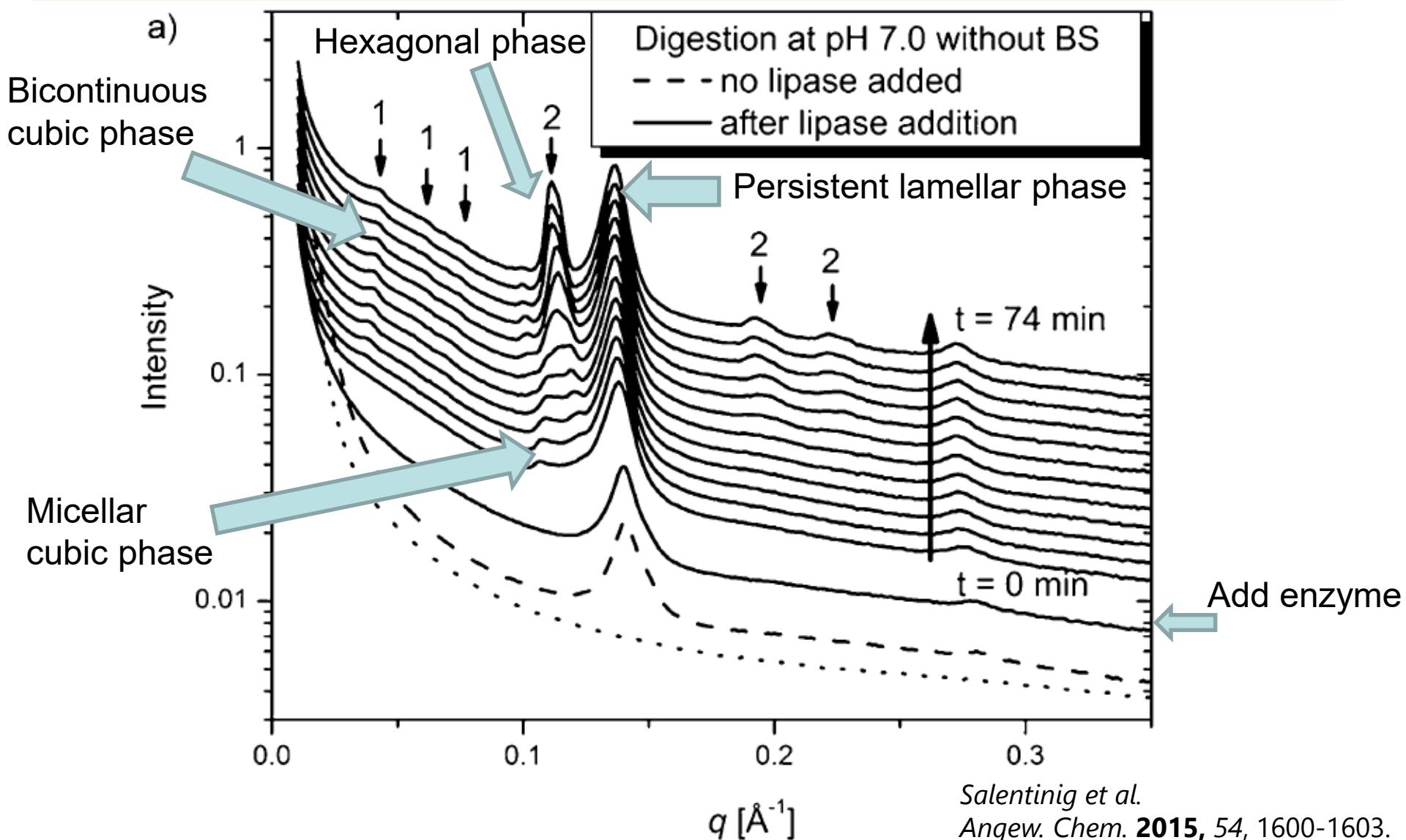
ORIGINAL RESEARCH
published: 05 December 2019
doi: 10.3389/fbioe.2019.00384

Internal Lamellar and Inverse Hexagonal Liquid Crystalline Phases During the Digestion of Krill and Astaxanthin Oil-in-Water Emulsions

Anan Yaghmur^{1*}, Saleh Lotfi^{1†}, Sarah Atoussa Ariabod^{1†}, Gizem Bor¹, Mark Gontsarik² and Stefan Salentinig^{2,3*}

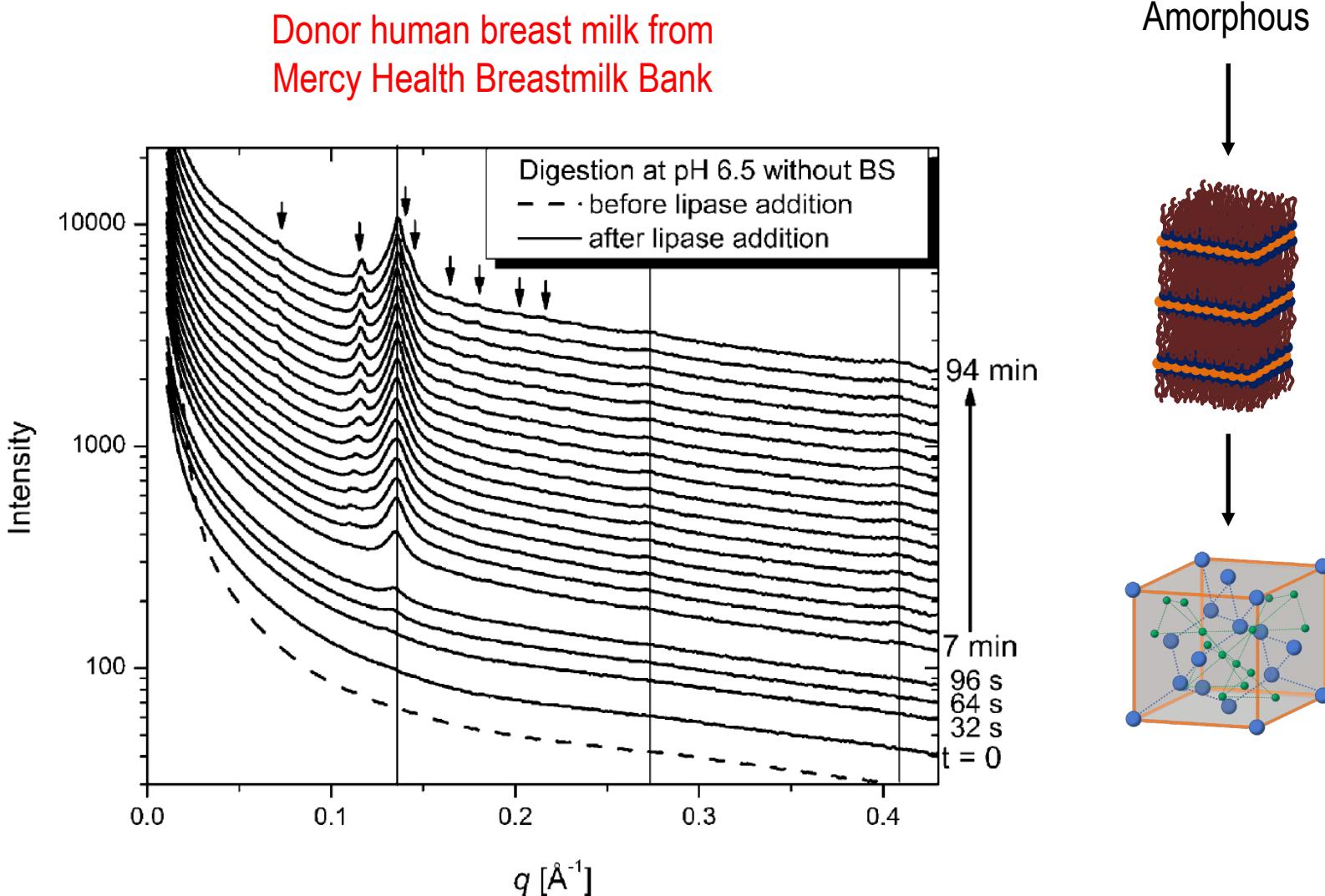


What do we 'see' when we digest human breast milk?



- Almost identical structural transitions as bovine milk
- Human breast milk 'self-digests' at 37°C due to bile salt-stimulated lipase

pH control is very important



- Inverted micellar cubic phase (I_2) main non-lamellar structure

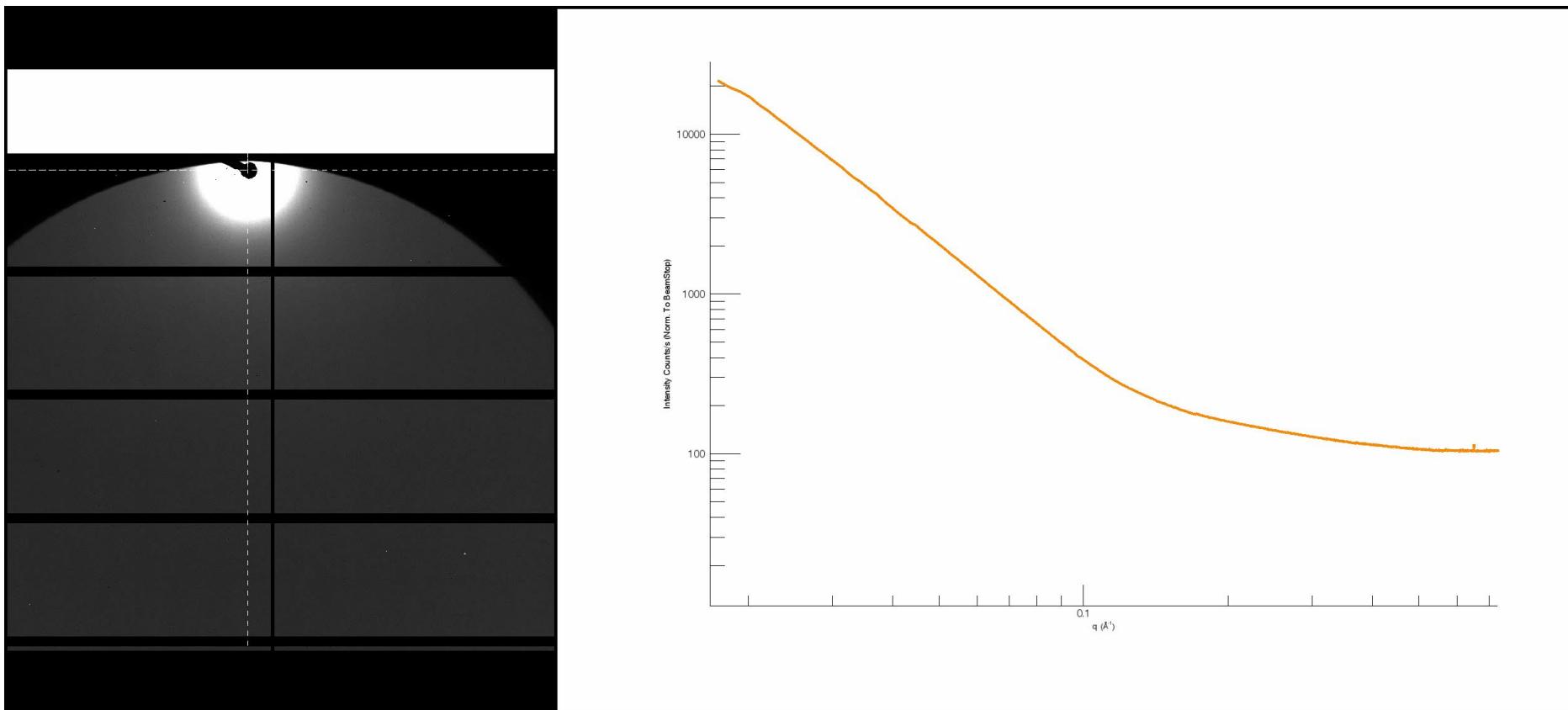
Published in Salentinig et al. *Angewandte Chemie* (2015), 54, 1600-1603.

Rare footage of the first wild almond to ever be milked (circa 1885)

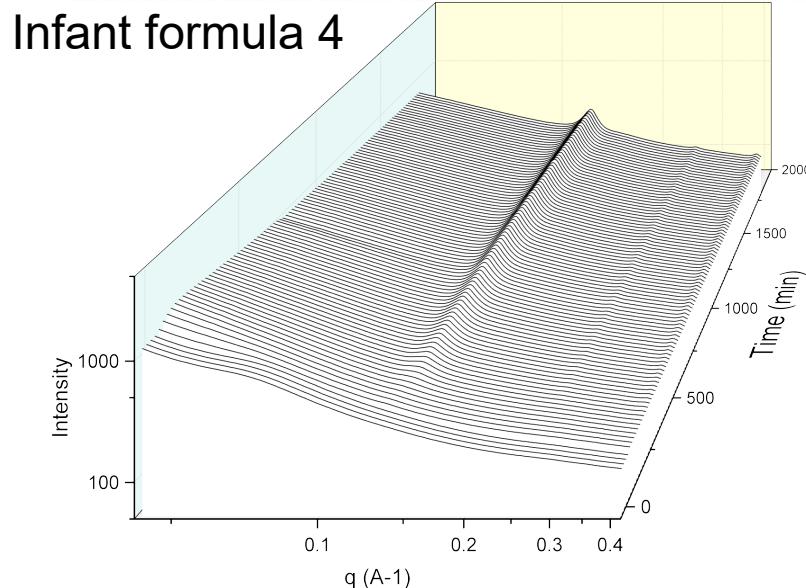
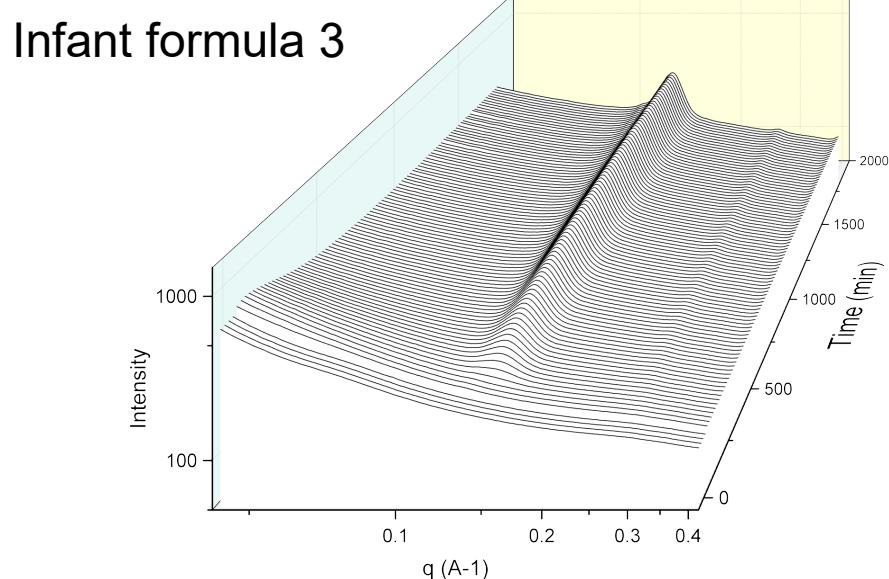
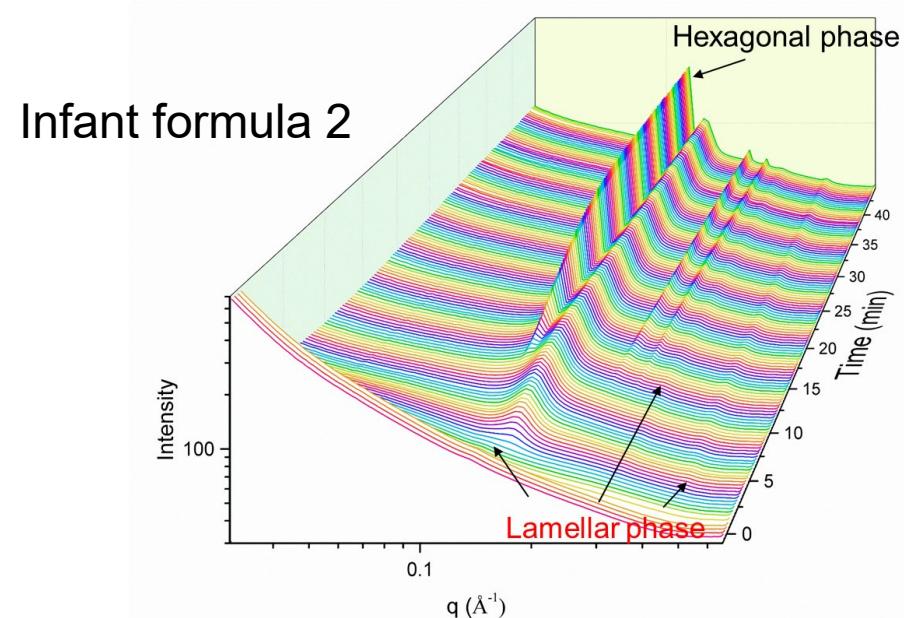
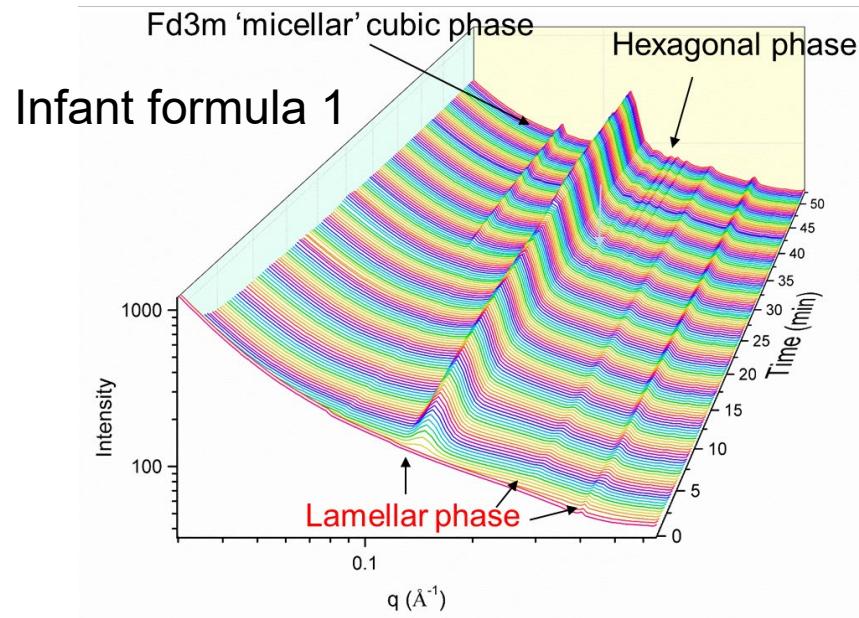


What do we 'see' with other 'so-called' milks?

Digestion of soy milk



Infant formula can be similar, or VERY different to milk

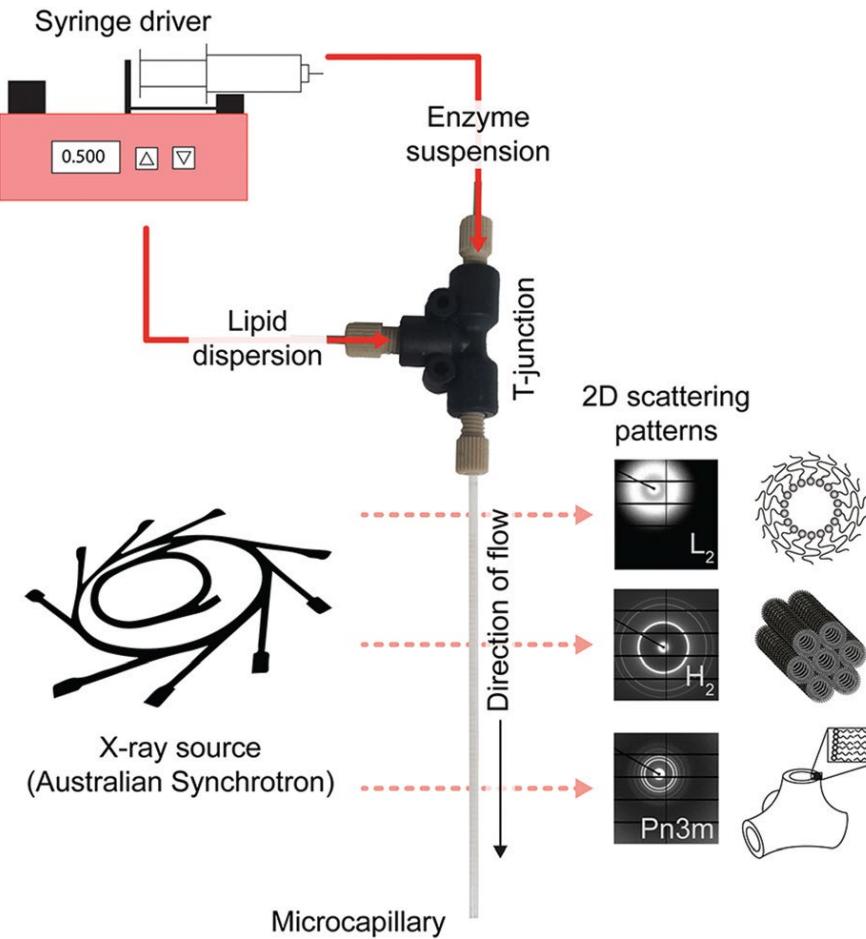


Lipid digestion and microfluidics

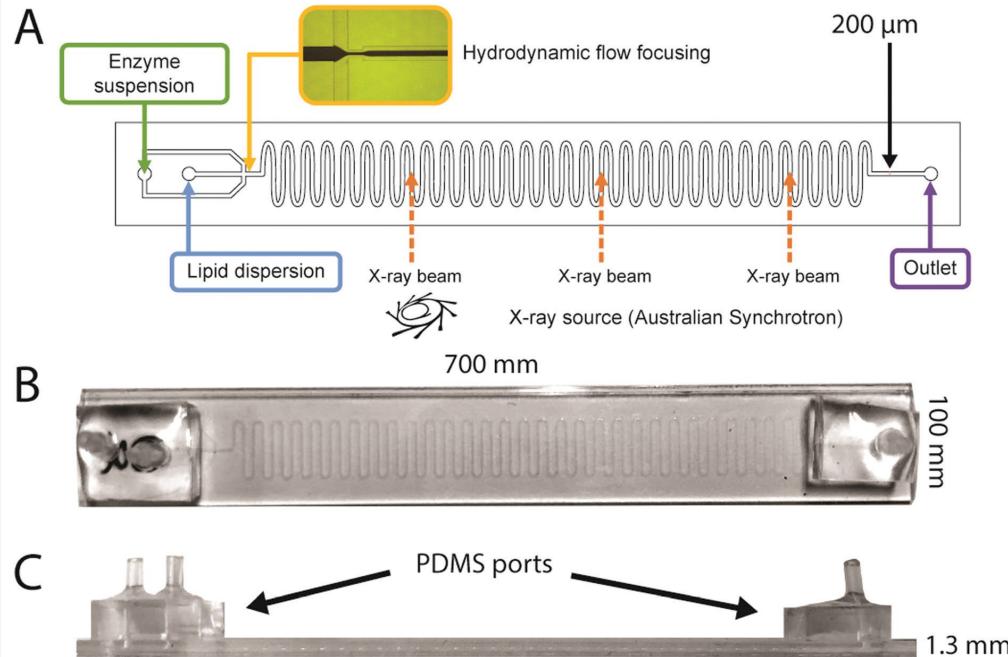
Linda Hong....Ben J. Boyd, Soft Matter, 2019



T-mixer



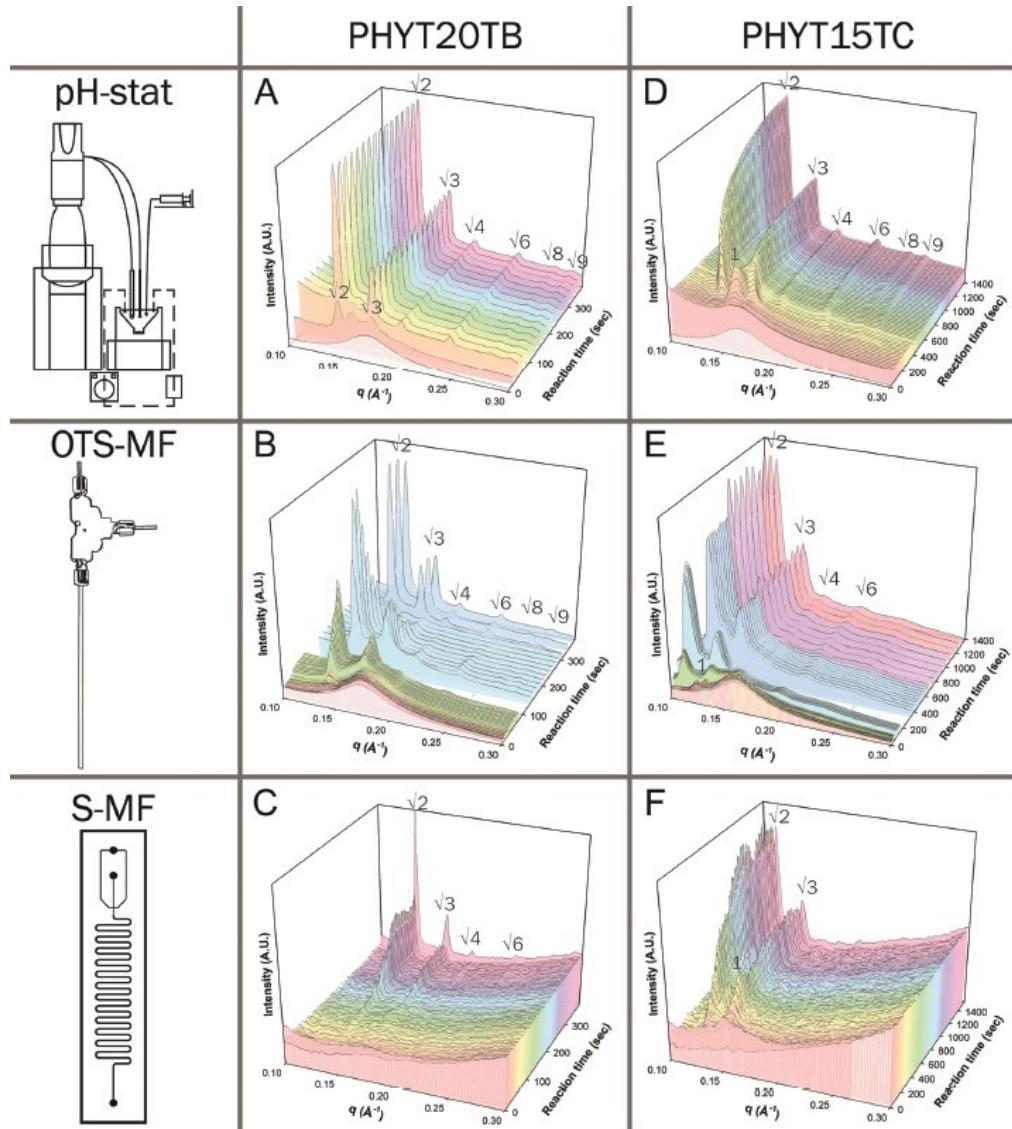
'Serpentine' flow focussing



Lipid digestion and microfluidics

Linda Hong....Ben J. Boyd, Soft Matter, 2019

- High throughput
- Structural changes in agreement
- Avoid radiation damage
- Spatial sampling of time
- Coupling to other techniques
- **Transmission can be an issue**



Structure from carbohydrates – maize starch

J. Blazek, E.P. Gilbert / Carbohydrate Polymers 85 (2011) 281–293



Carbohydrate Polymers 85 (2011) 281–293

Contents lists available at ScienceDirect

Carbohydrate Polymers

journal homepage: www.elsevier.com/locate/carpol

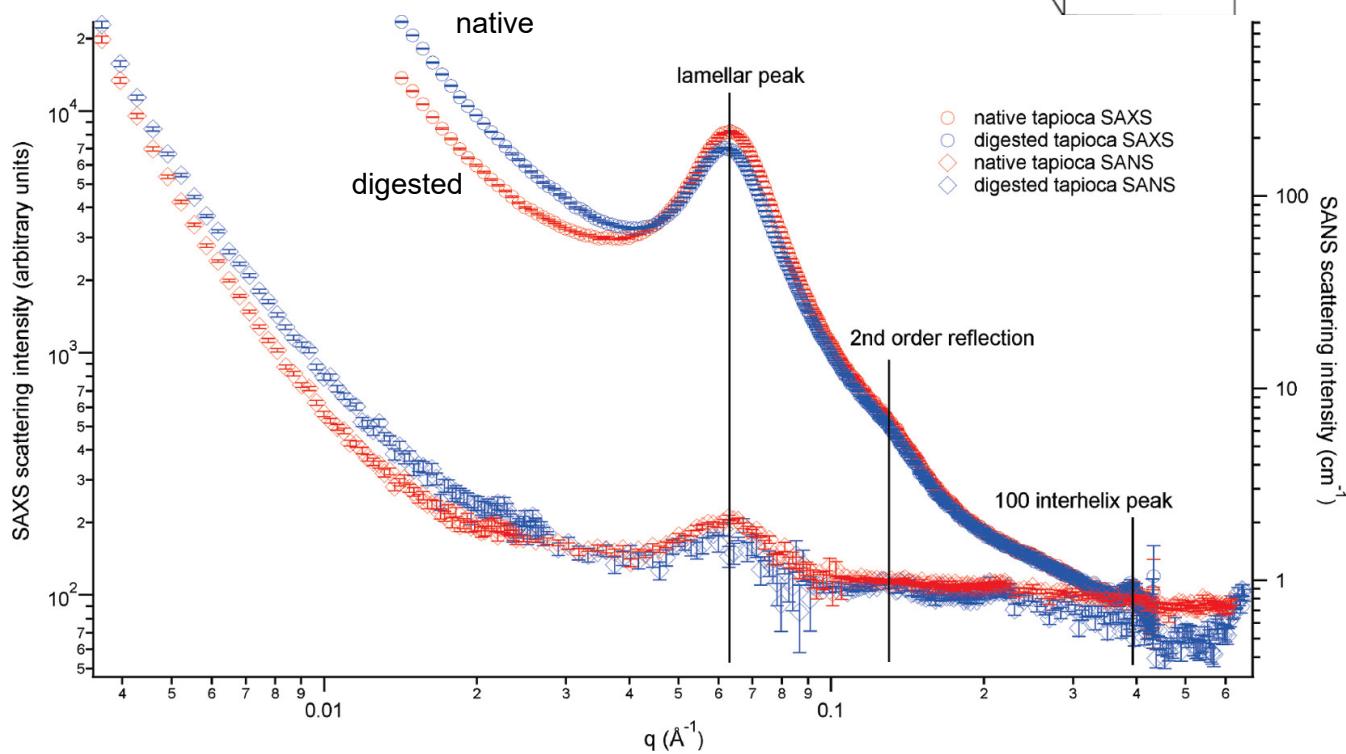
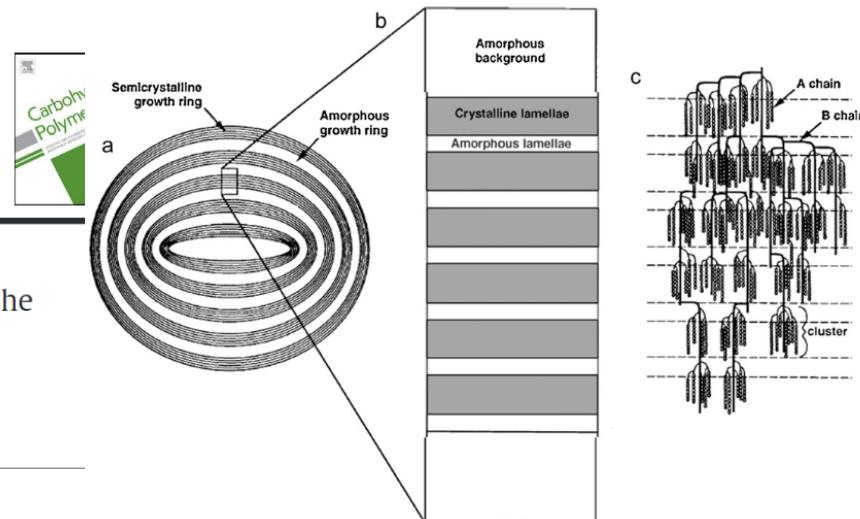
Review

Application of small-angle X-ray and neutron scattering techniques to the characterisation of starch structure: A review

Jaroslav Blazek^{a,b}, Elliot Paul Gilbert^{a,*}

^a Bragg Institute, Australian Nuclear Science and Technology Organisation, Locked Bag 2001, Kirrawee DC, NSW 2232, Australia

^b Food Futures Flagship, North Ryde Riverside Corporate Park, 5 Julius Avenue, North Ryde, NSW 2113, Australia



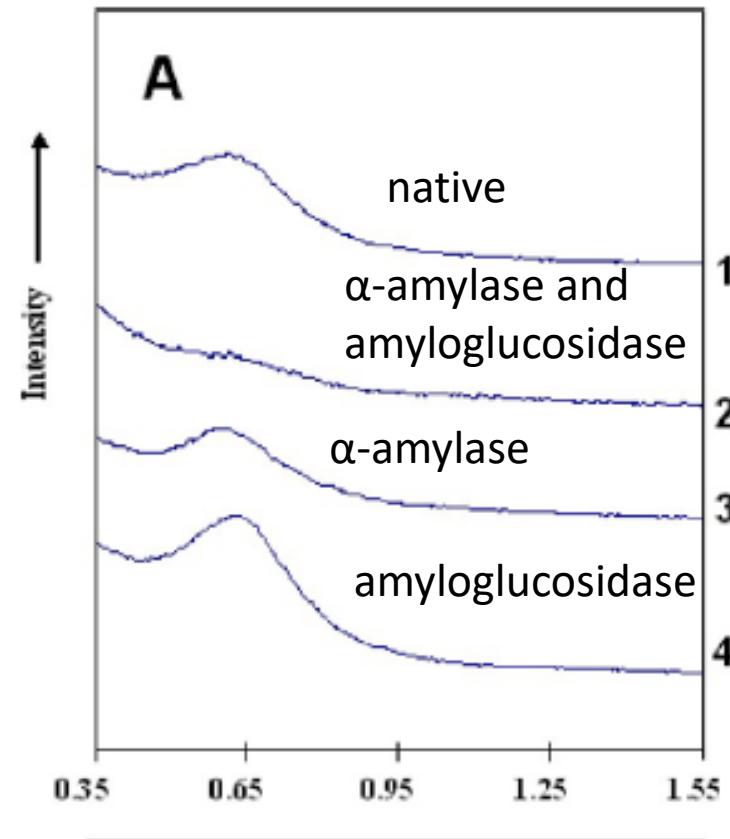
Structure from carbohydrates – maize starch

Mechanism and Enzymatic Contribution to In Vitro Test Method of Digestion for Maize Starches Differing in Amylose Content

Lauren R. Brewer, Liming Cai, and Yong-Cheng Shi*

College of Agriculture, Department of Grain Science and Industry, Kansas State University, Manhattan, Kansas 66506, United States

- 'Before and after' study
- Incubated with enzymes for 120 min (I think)



- 9 nm lamellar peak at $q \sim 0.65$ ($1/\text{nm}$) attributable to the alternative repeating stacks of amorphous and crystalline lamella in starch granules.
- Enzyme mix synergistic in degrading amorphous and crystalline regions (based also on e.g. HPLC analysis)

Structure from carbohydrates – resistant starch

Biomacromolecules 2008, 9, 1951–1958

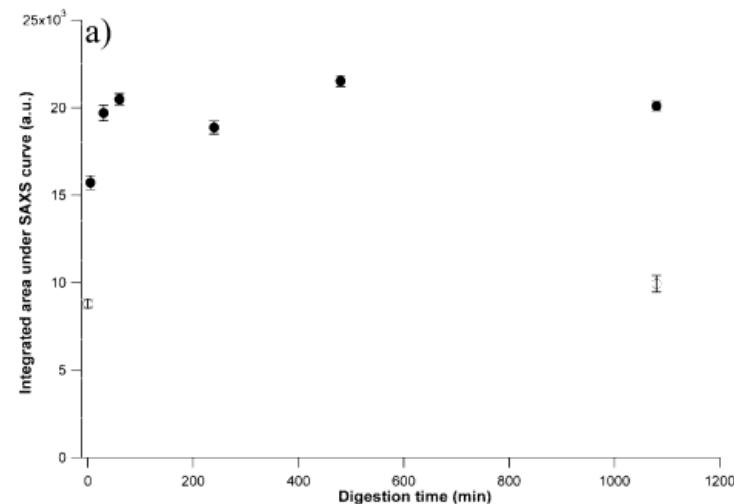
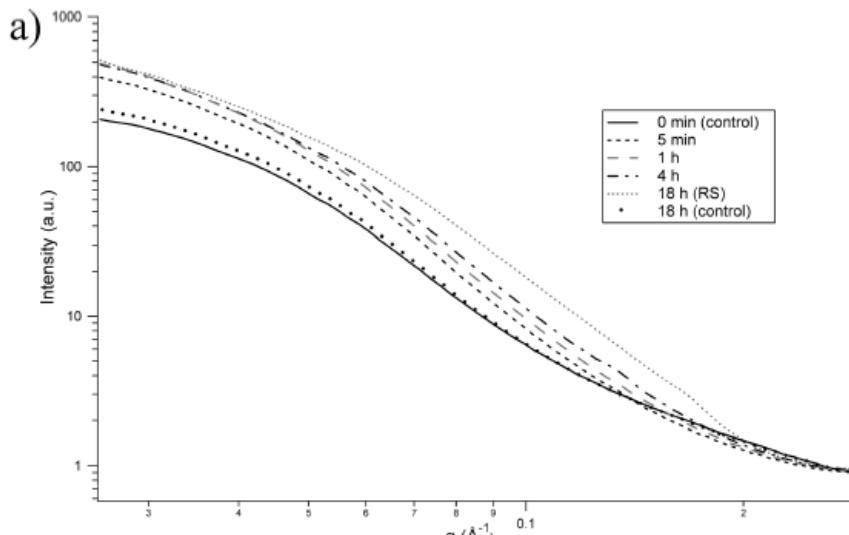
1951

Molecular Rearrangement Of Starch During In Vitro Digestion: Toward A Better Understanding Of Enzyme Resistant Starch Formation In Processed Starches

Amparo Lopez-Rubio,^{*,†} Bernadine M. Flanagan,[‡] Ashok K. Shrestha,^{‡,§}
Michael J. Gidley,[‡] and Elliot P. Gilbert[†]

1954 *Biomacromolecules*, Vol. 9, No. 7, 2008

Lopez-Rubio et al.



processing conditions as described elsewhere.^{12,13} (1) “Mild” conditions → 50% moisture content, 100 °C and 150 s⁻¹ shear rate; (2) “Extreme” conditions → 35% moisture content, 140 °C and 750 s⁻¹ shear rate.

After processing, the samples were freeze-dried to avoid retrogradation and digested according to the method described in refs 12 and 13.

Isolation of RS Fractions at Different Stages of Digestion. Samples were collected at different times from the digestion solution. Alcohol was added to inactivate the enzymes and the samples were immediately freeze-dried to avoid structural modifications. Two control samples were also prepared in which the extruded samples were kept in the same buffer used for digestion, but without starch hydrolyzing enzymes: Control $t = 0$ (the extruded samples were added into the buffer at 37 °C in which alcohol was added and the samples immediately freeze-dried) and control $t = 18$ h (the extruded starches were kept in the buffer at 37 °C for 18 h in a shaking water bath; after 18 h, alcohol was added and the samples were freeze-dried).

Small Angle X-ray Scattering (SAXS). SAXS measurements were performed on a Bruker Nanostar SAXS camera, as described previously.¹³

Structure from carbohydrates – corn starch

International Journal of Biological Macromolecules 129 (2019) 361–369



Contents lists available at ScienceDirect

International Journal of Biological Macromolecules

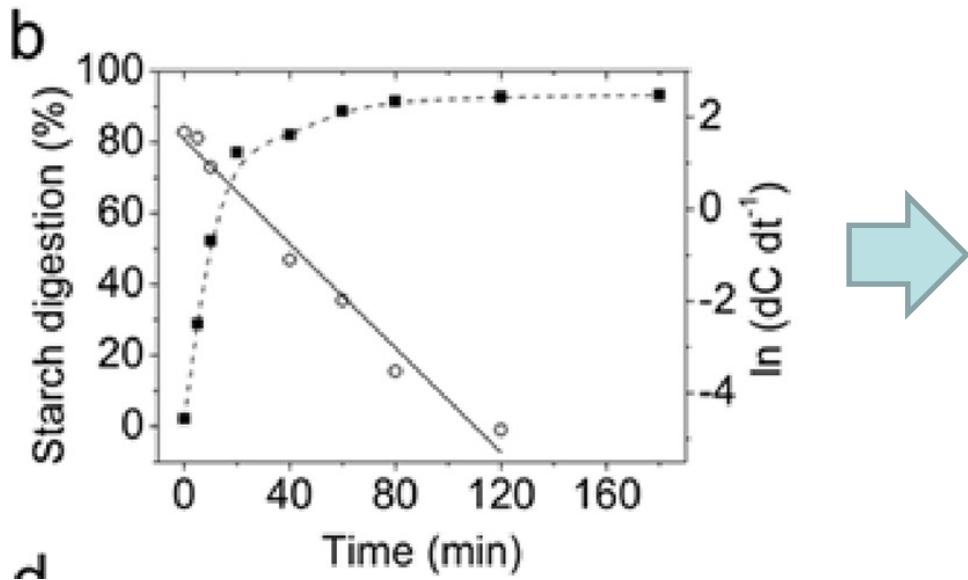
journal homepage: <http://www.elsevier.com/locate/ijbiomac>



Structurally induced modulation of in vitro digestibility of amylopectin corn starch upon esterification with folic acid



Pallab Kumar Borah ^{a,b}, Michael Rappolt ^a, Raj Kumar Duary ^{b,*}, Anwesha Sarkar ^{a,*}

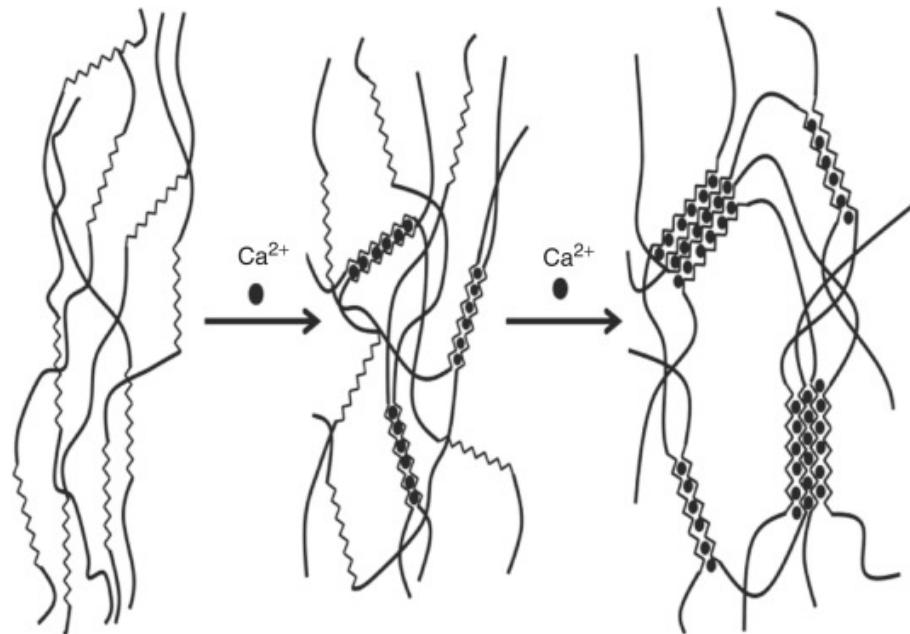


Time resolved diffraction?

Gastric pH 2.2, with pepsin

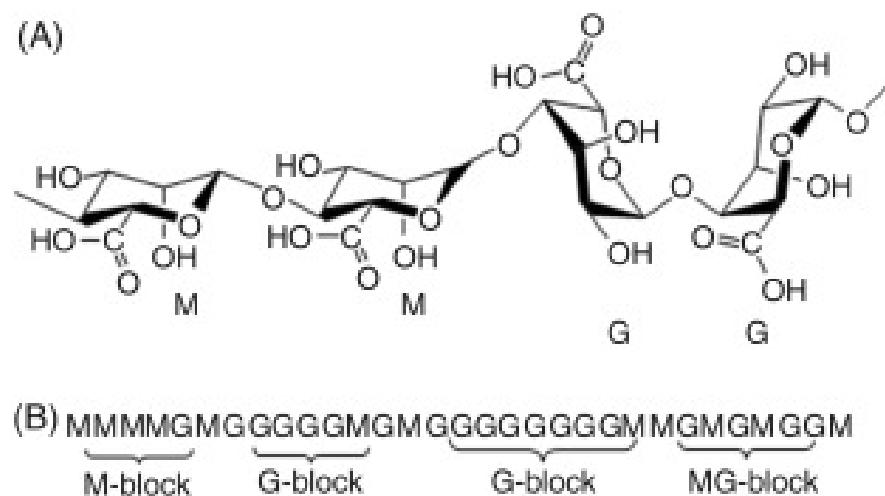
Intestinal pH 6.8 pancreatin, amylase, protease, amyloglucosidase and invertase

Structure from carbohydrates - Alginate gels



From brown algae
Calcium cross-linked
Properties depend on M/G ratio
"Egg box" model

(1 \rightarrow 4)-linked β -D -mannuronic acid (M) and α -L-gluronic acid (G)



Structure from carbohydrates - Alginate gels

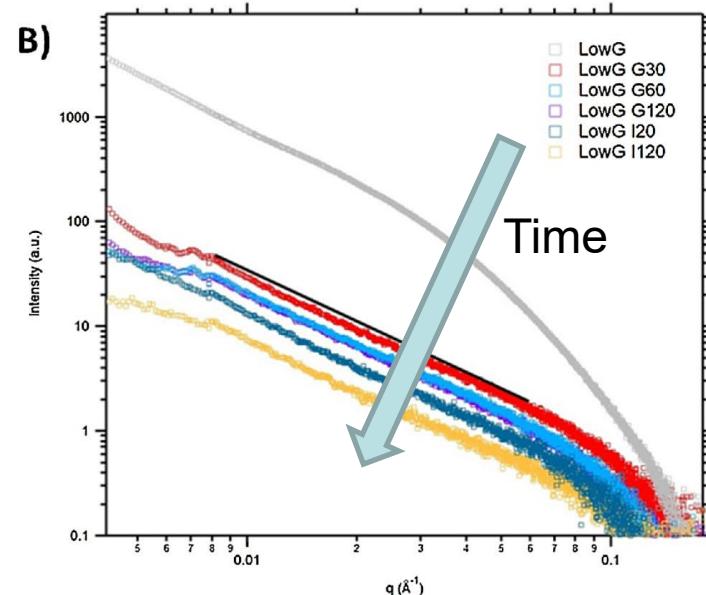
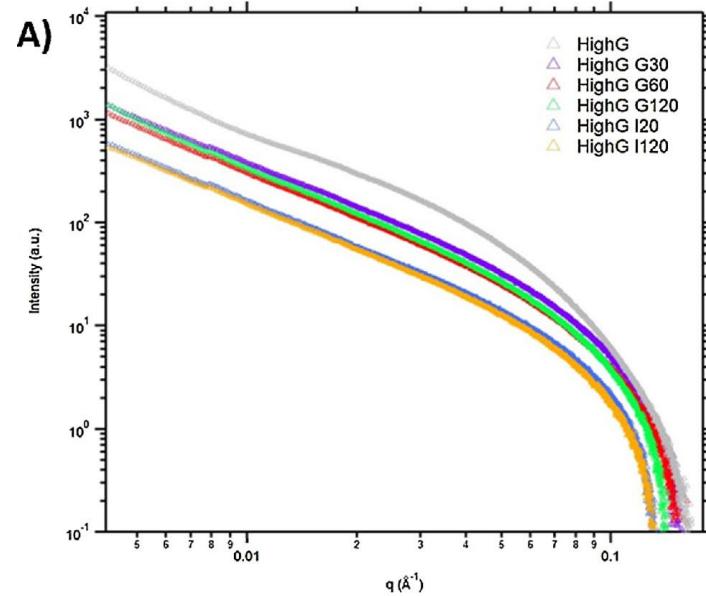
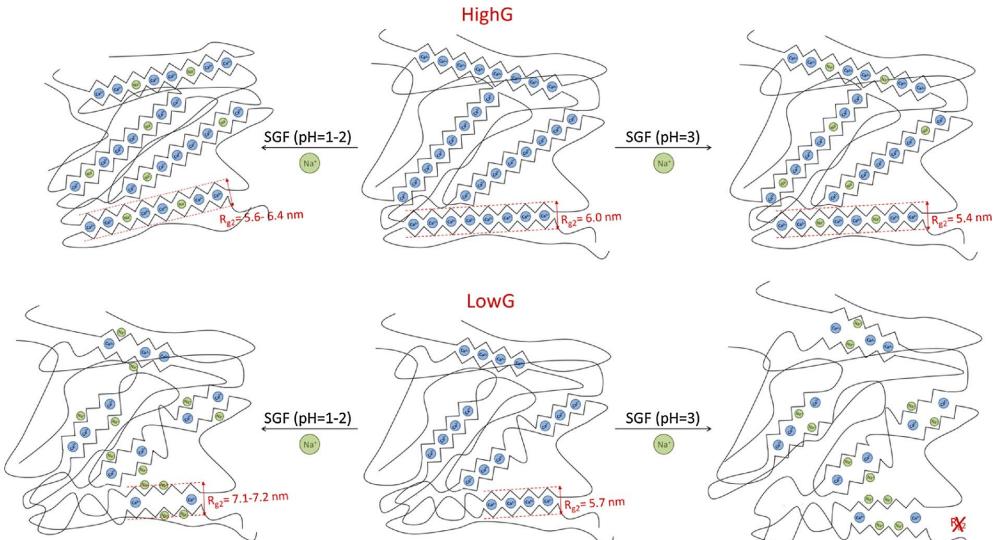
ABSTRACT

Alginate microcapsules were prepared using three different alginate grades and incubated under simulated digestion conditions

3.3. Impact of *in vitro* gastrointestinal digestion on the alginate microbeads

In order to assess the micro- and nanostructural changes undergone by the alginate microcapsules upon digestion, these were subjected to simulated gastrointestinal conditions, as described in the standardised static *in vitro* digestion method, developed within the INFOGEST international network (Brodkorb et al., 2019; Minekus et al., 2014). Accordingly, the microcapsules were incubated in simulated gastric fluid (SGF) at a constant pH of 3, and simulated intestinal fluid (SIF, pH = 7), consecutively, at 37 °C for 2 h periods. However, no digestive enzymes or bile were added in this study, since their presence could give rise to the appearance of scattering features, interfering with the

Ca^{2+} apparently displaced by Na^+



Structure from proteins

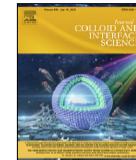
Journal of Colloid and Interface Science 594 (2021) 561–574



Contents lists available at ScienceDirect

Journal of Colloid and Interface Science

journal homepage: www.elsevier.com/locate/jcis



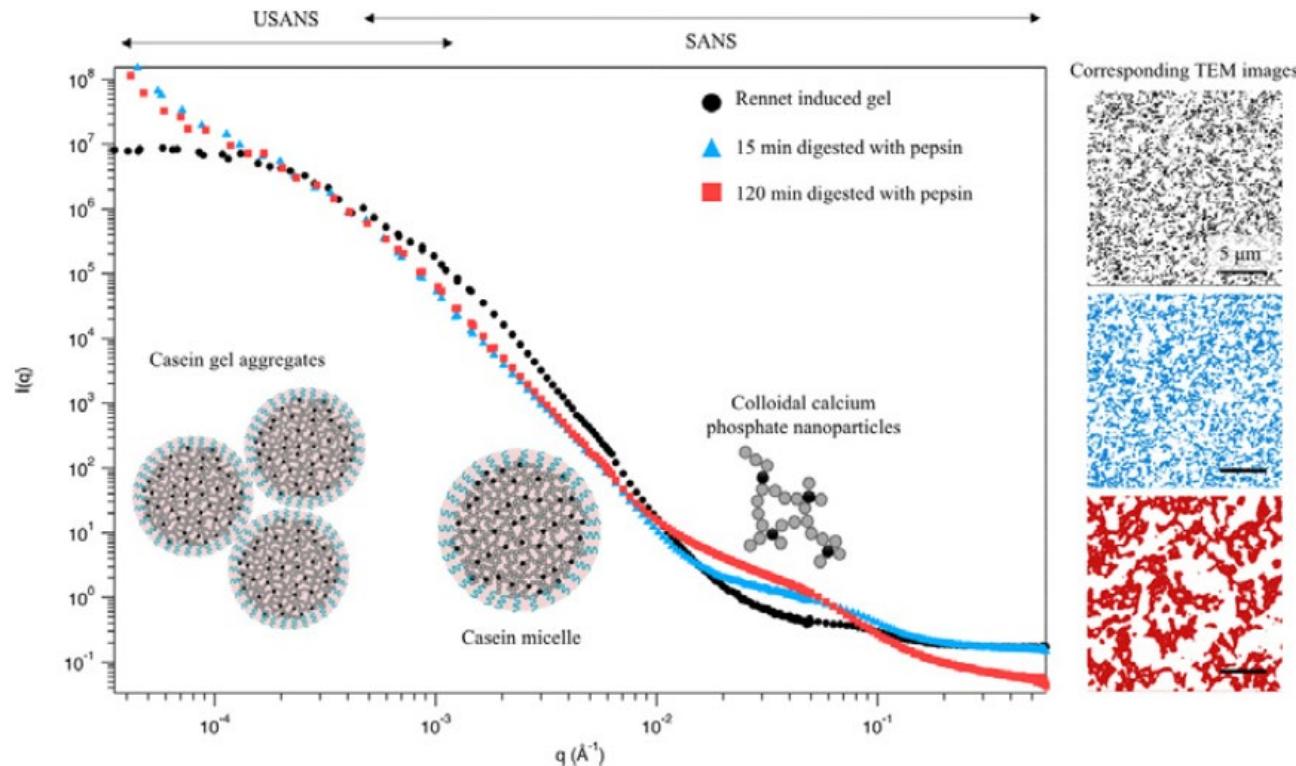
Regular Article

Investigating casein gel structure during gastric digestion using ultra-small and small-angle neutron scattering



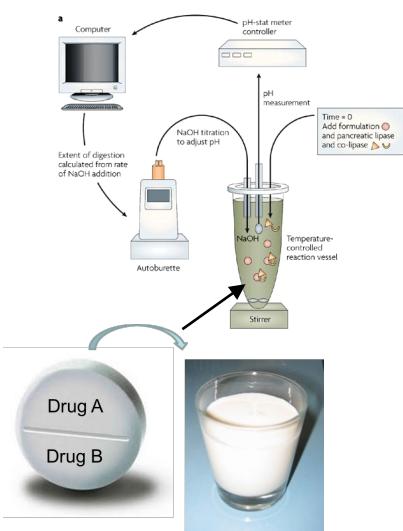
Meltem Bayrak ^{a,b,*}, Jitendra Mata ^c, Jared K. Raynes ^a, Mark Greaves ^d, Jacinta White ^d, Charlotte E. Conn ^b,
Juliane Flory ^e, Amy Logan ^a

Lots of studies using scattering on casein micelles
Very few actually during digestion

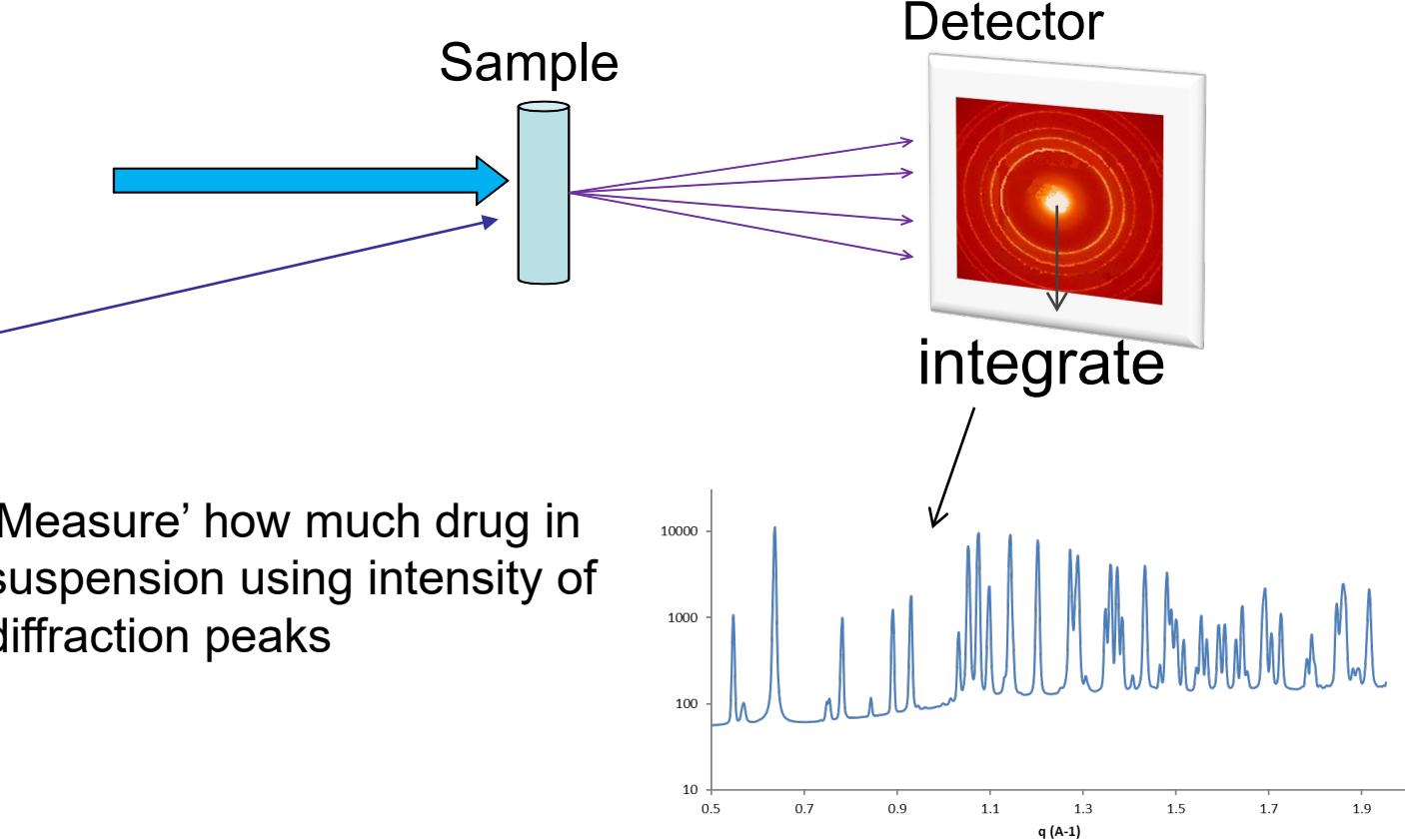


In situ determination of drug disposition

Synchrotron X-ray source

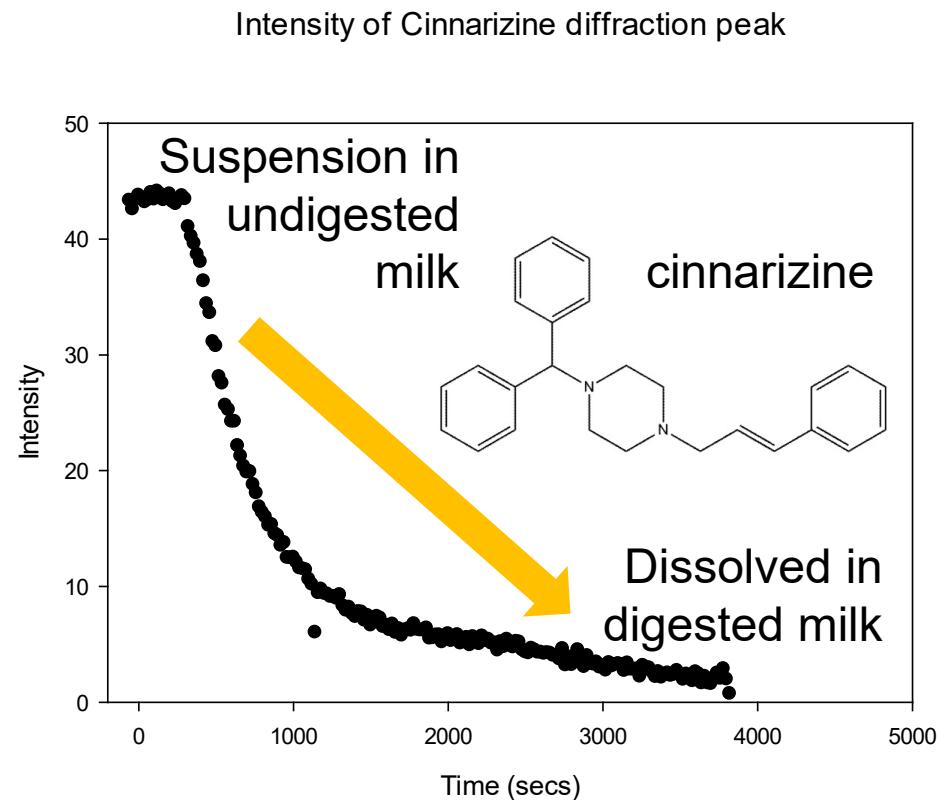
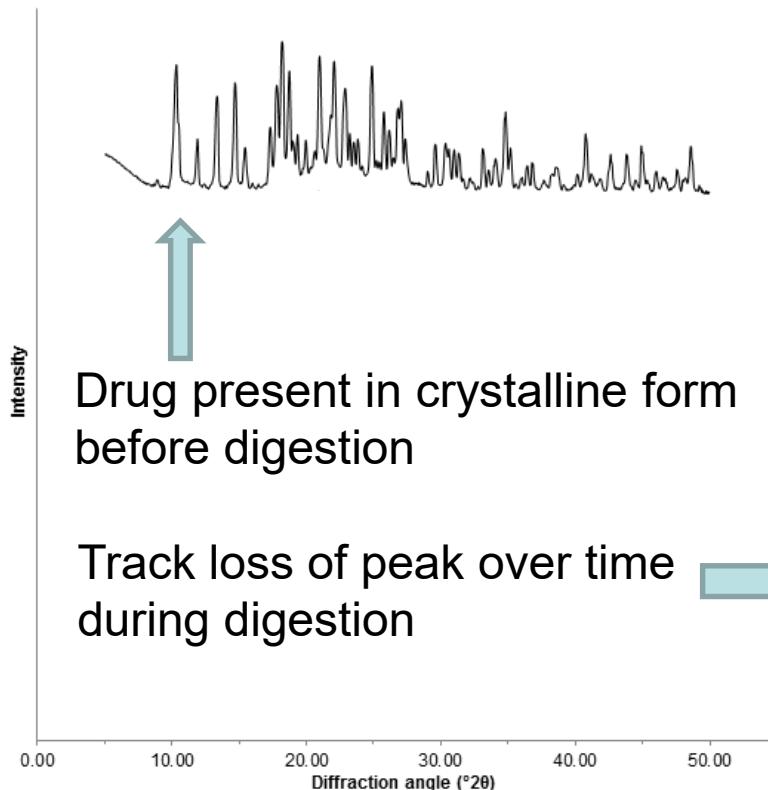


'Measure' how much drug in suspension using intensity of diffraction peaks



- 'Sees' crystalline drug through lipid emulsion avoiding 'separate and sample' difficulties esp. with milk
 1. **Appearance of peaks = precipitation**
 2. **Disappearance = solubilisation/amorphization**
 3. **Change = polymorphic transformation**

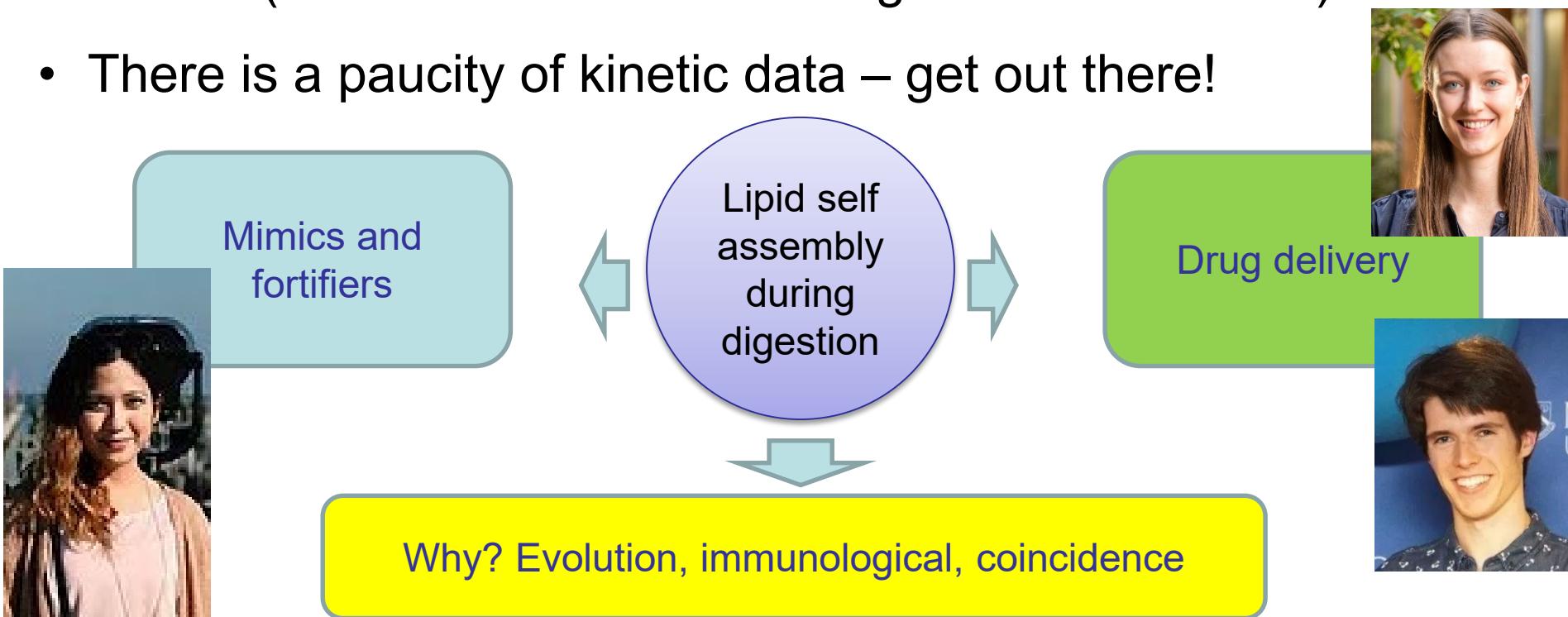
Real-time solubilization of drug during digestion of milk



- Drug has low solubility in milk – drug from disintegrating tablet would not dissolve in primarily milk medium
- Drug is completely soluble in ‘digested’ milk
- Correlates with oral bioavailability

Take home points

- Structure is important – especially during and post-digestion
- Scattering can yield new understanding in food and other soft matter in physiological conditions especially using synchrotron source (but beware need for background subtraction)
- There is a paucity of kinetic data – get out there!



Some practicalities

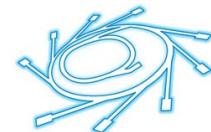
1. Beware radiation damage (in any *in situ* kinetic experiment)
2. Think ahead about need for background subtraction. Flow through is good but not bullet proof and often not 'real' background.
3. Autosampler coupled with flowthrough probably most ideal
4. Consider physiological reality with respect to complexity (which can make 2. difficult), volumes, and also time. Enzymes are usually extracts = high and variable background which in turn makes modelling difficult....
5. Imagination gets you a long way, talking to your beamline staff beforehand gets you even further.

Acknowledgements



@group_boyd

nonlamellar.com



Australian Synchrotron
Turning bright ideas into brilliant outcomes

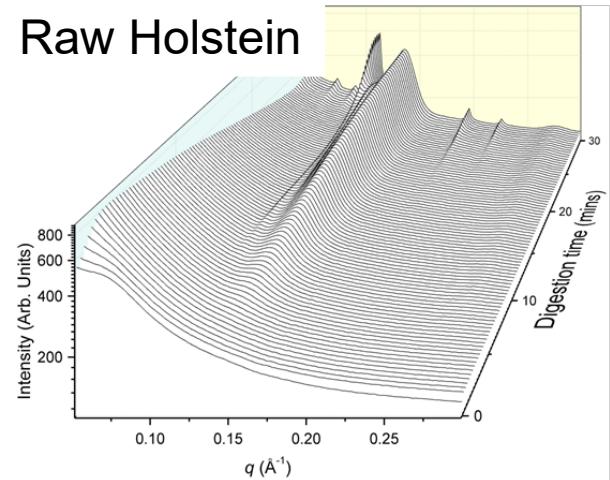


Australian Government
Australian Research Council

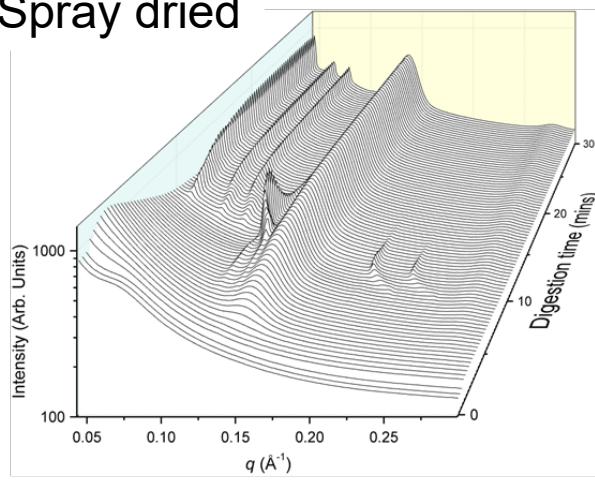
BILL & MELINDA
GATES foundation

What can we do to milk to upset its behaviour?

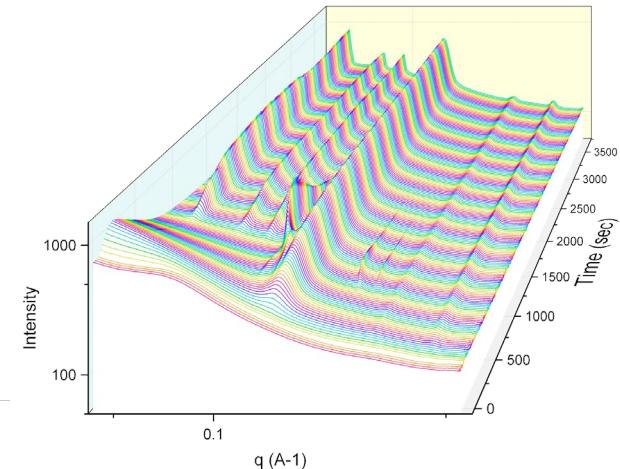
Raw Holstein



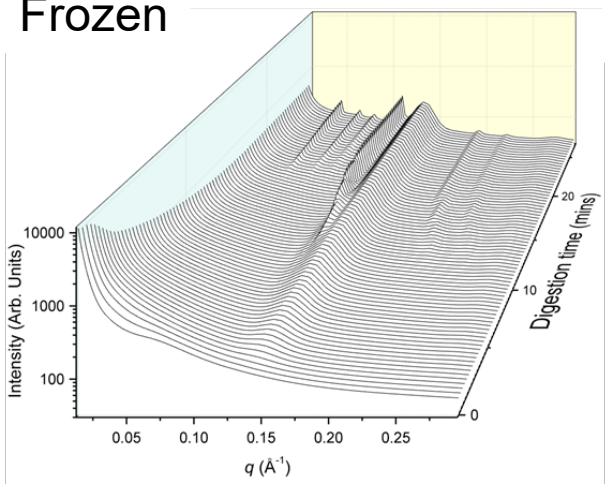
Spray dried



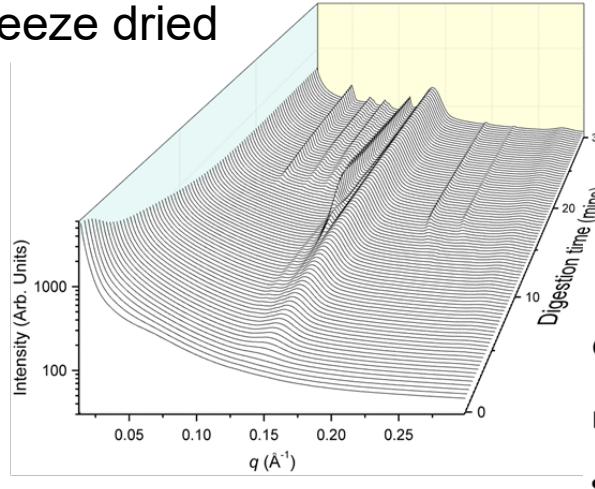
Commercial



Frozen

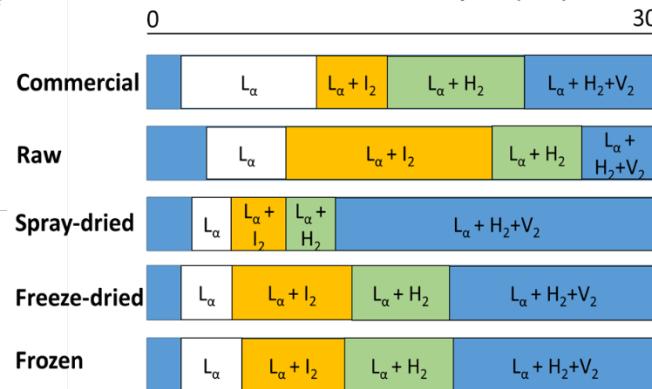


Freeze dried



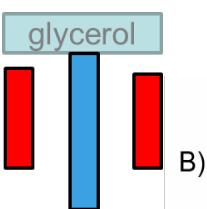
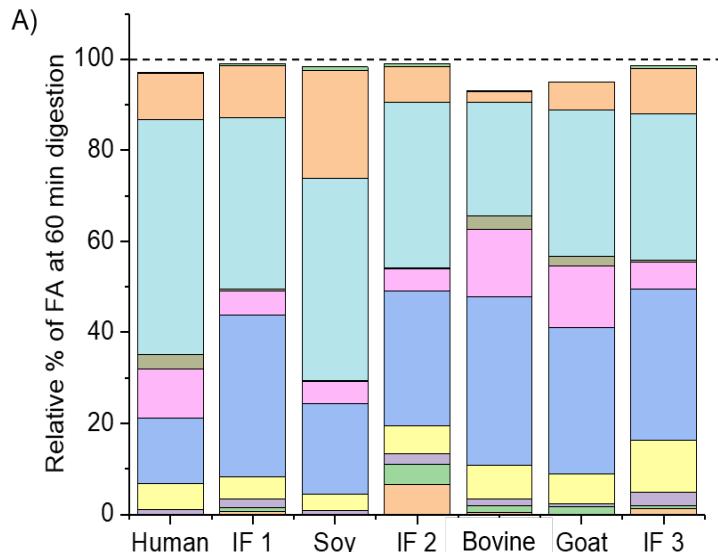
- It's reasonably difficult!

Time after addition of lipase (min)

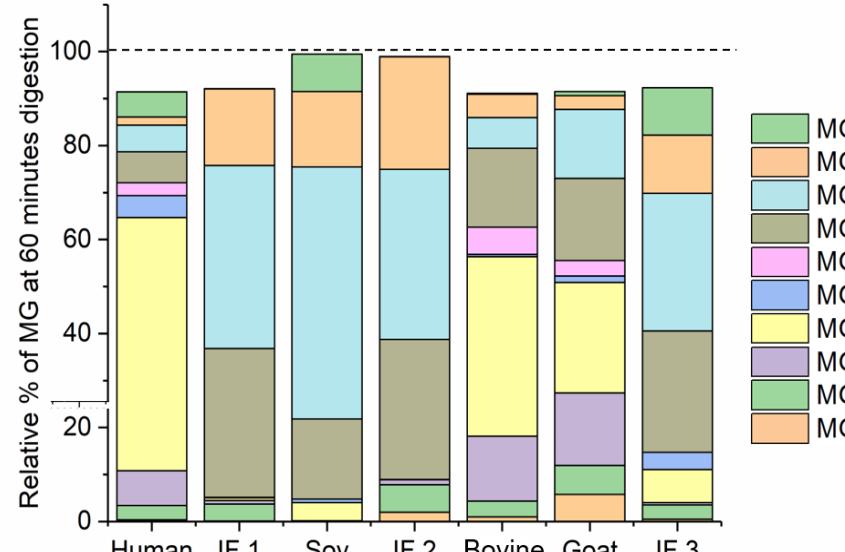


Lipid distribution after digestion complete

Free fatty acid distribution



Monoglyceride distribution



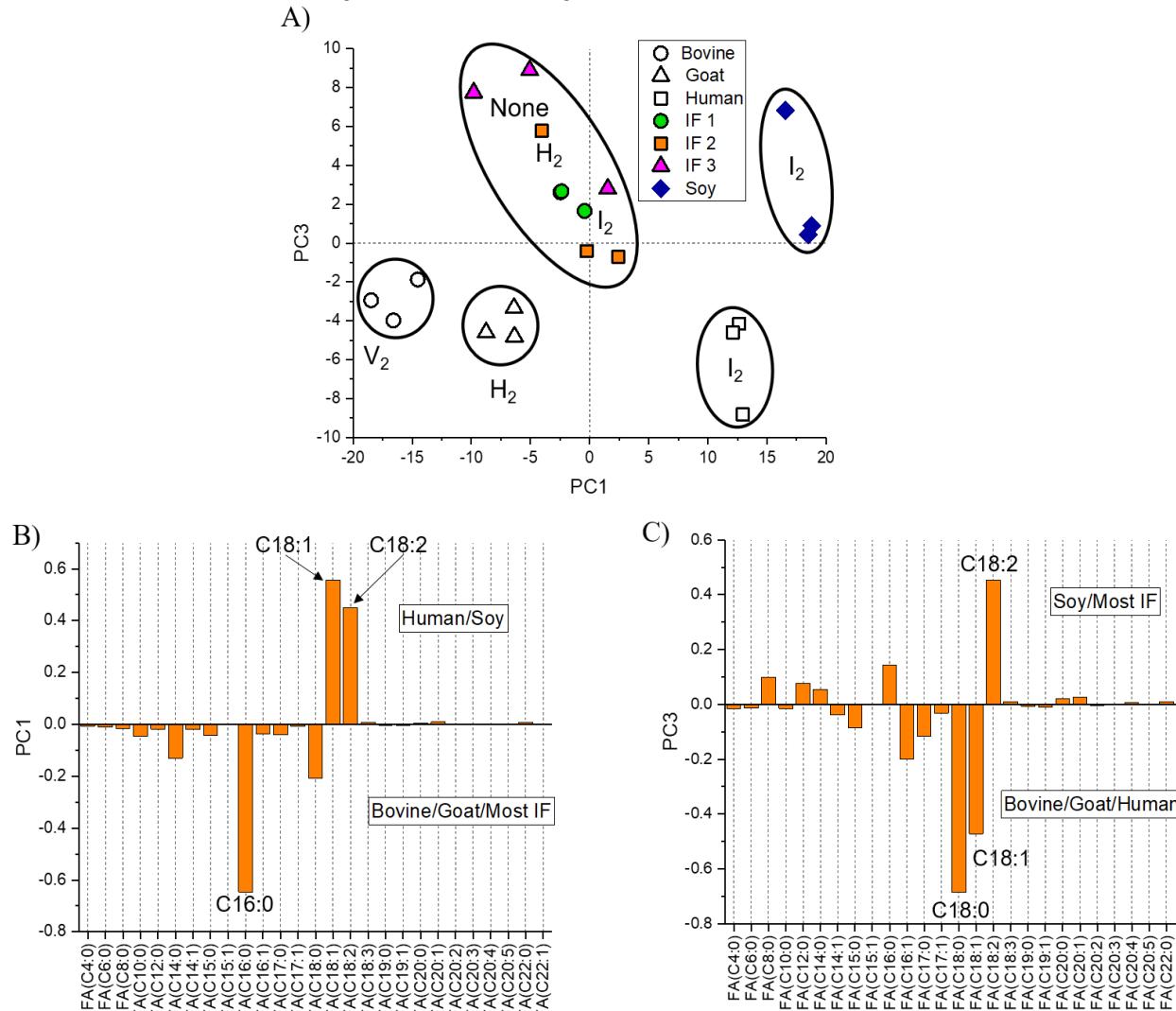
LC structures	I ₂	I ₂	I ₂	H ₂	V ₂	L _α	L _α
	L _α	L _α	L _α	L _α	H ₂	L _α	L _α

LC structures	I ₂	I ₂	I ₂	H ₂	V ₂	L _α	L _α
	L _α	L _α	L _α	L _α	H ₂	L _α	L _α

- Monopalmitin much more prevalent in mammal milks than others
- Free fatty acids not as variable
- Infant formulas more greatly resemble Soy (+ other vegetable oils) than milk

Lipid distribution after digestion complete

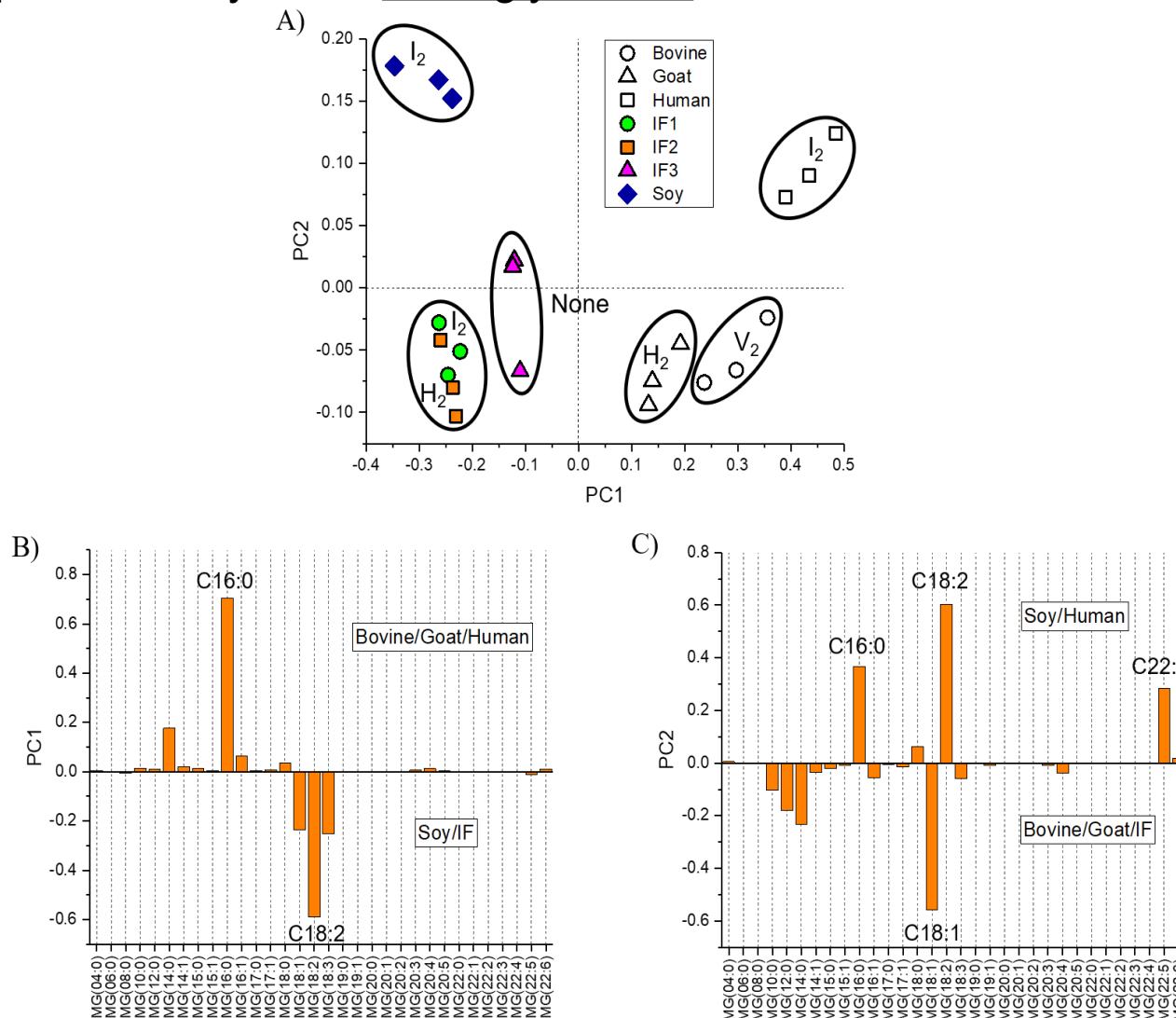
Principal component analysis for fatty acids



- Primary differences are driven by high oleic/linoleic acid, low palmitic acid

Lipid distribution after digestion complete

Principal component analysis for monoglycerides



- Primary differences are driven by high monopalmitin, low monoolein/linolinolein