



# LINX Membrane Protein Workshop

25<sup>th</sup> – 26<sup>th</sup> May, 2021



EUROPEAN  
SPALLATION  
SOURCE

## Towards Neutron Crystallography: Insights into production of deuterium-labelled OmpF

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Supervisor: Dr. Esko Oksanen



Proton



Electron

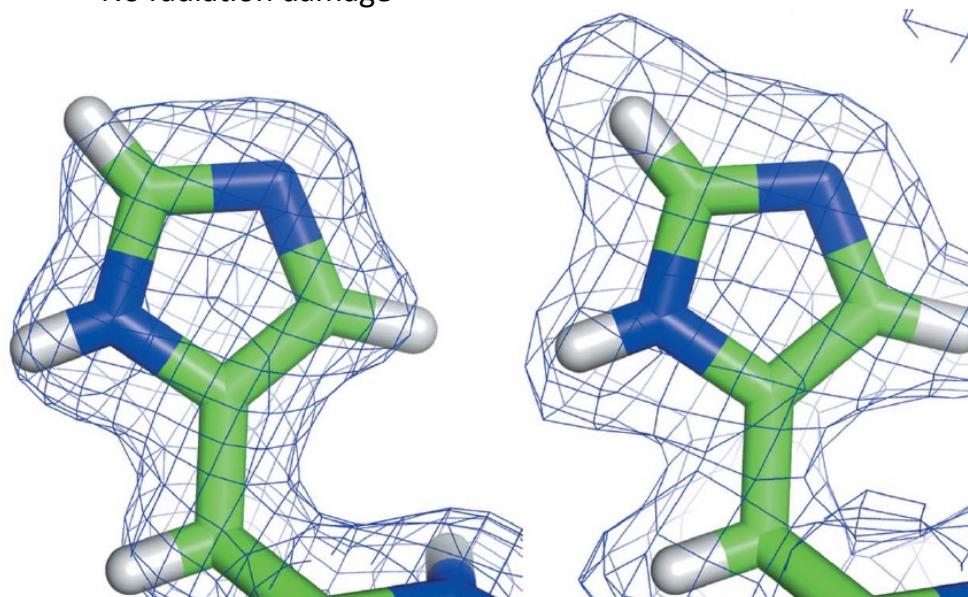


Neutron

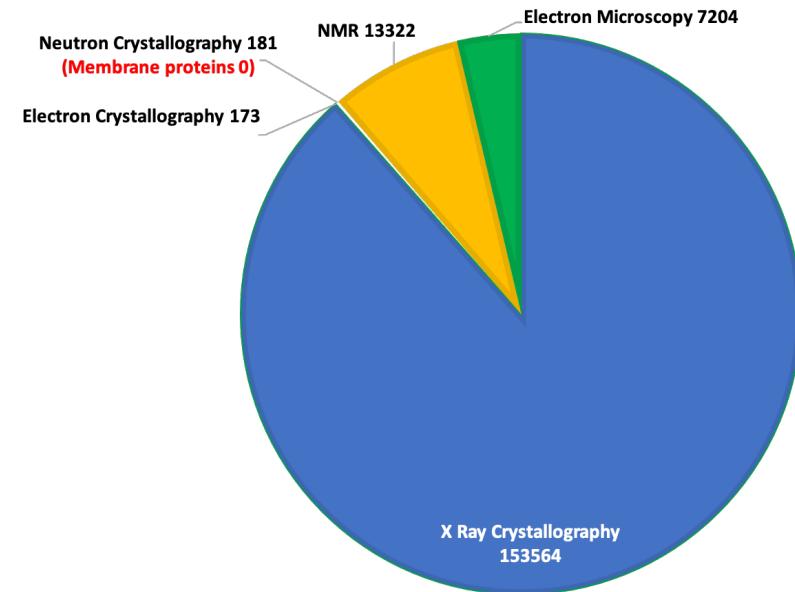


# Importance of Neutron crystallography

- Neutrons interact with nuclei and independent of atomic number
- Hydrogens maybe visible even when the crystals diffracting X-rays to (sub-)atomic resolution
- Understand the protonated state of chemical groups, orientation of methyl and hydroxyl groups
- Data collection can be performed at room temperature
- No radiation damage



X-ray (left) and neutron (right) 2mFo-DFc maps (contoured at = 1.0) of His187 in the TIM structure. Kelpas et al, 2018



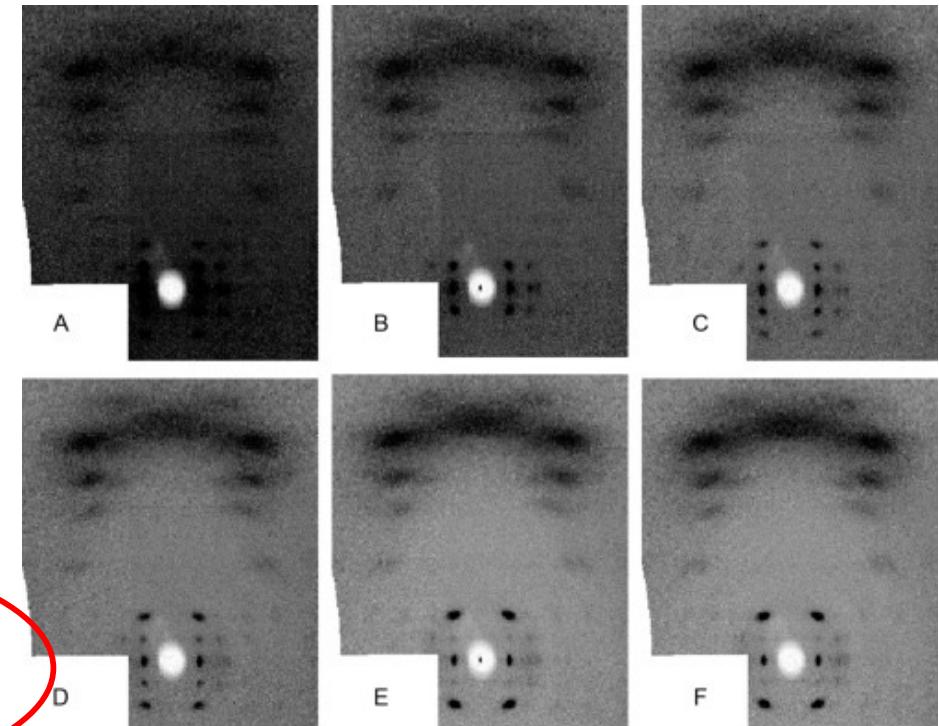
Reported protein structures in PDB obtained by using different experimental methods

**Bottlenecks:**  
 Weak flux of neutron beam  
 Large crystals  
 Negative coherent scattering of Hydrogen

# Purpose of D-labelling

- Incoherent scattering from hydrogen reduces the signal-to-noise ratio of the data
- Scattering length of deuterium is similar in sign and magnitude to those of C, N and O (Table 1).
- Deuterium should be as visible as carbon at 2.0 Å ° resolution.
- Perdeuterated crystals allow suitable neutron diffraction data at lower crystal volume (<1 mm<sup>3</sup>).

Atom	Atomic Number	Incoherent Cross-section(barn)	Neutron Scattering Length (fm)
Hydrogen	1	79.6	-3.74
Deuterium	1	2	6.67
Carbon	12	0	6.65
Nitrogen	14	0.3	9.3
Oxygen	16	0	5.8



Improvement in the visibility of the coherent diffraction peaks of A-DNA as the hydration of the sample is changed from H<sub>2</sub>O to D<sub>2</sub>O (Forsyth et al, 2016).

Table 1. Neutron scattering length and cross sections of elements.  
(Adapted from Sears, 1992)

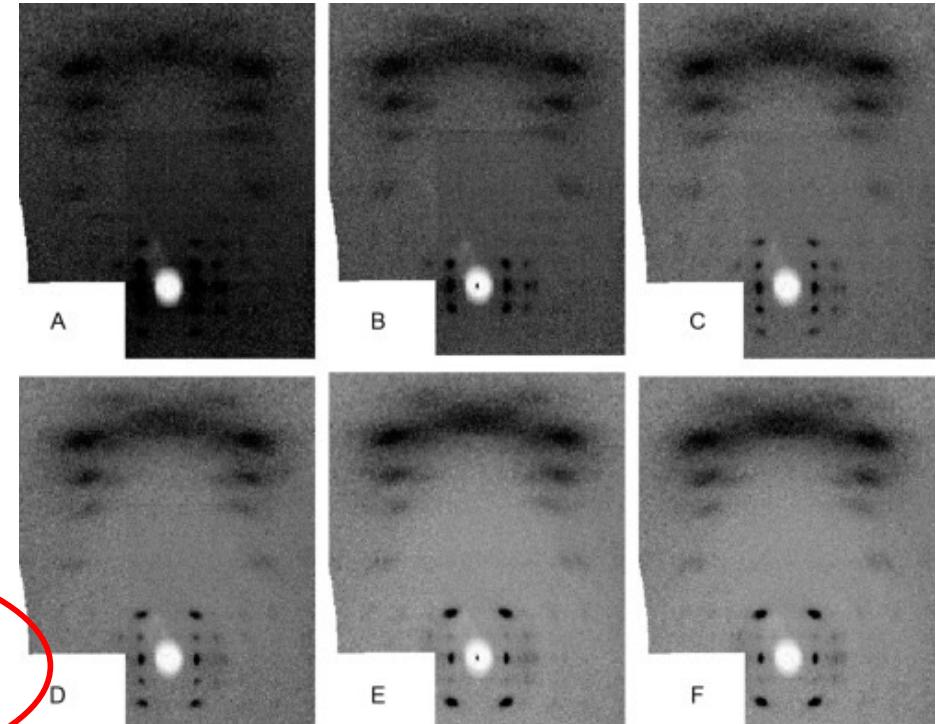
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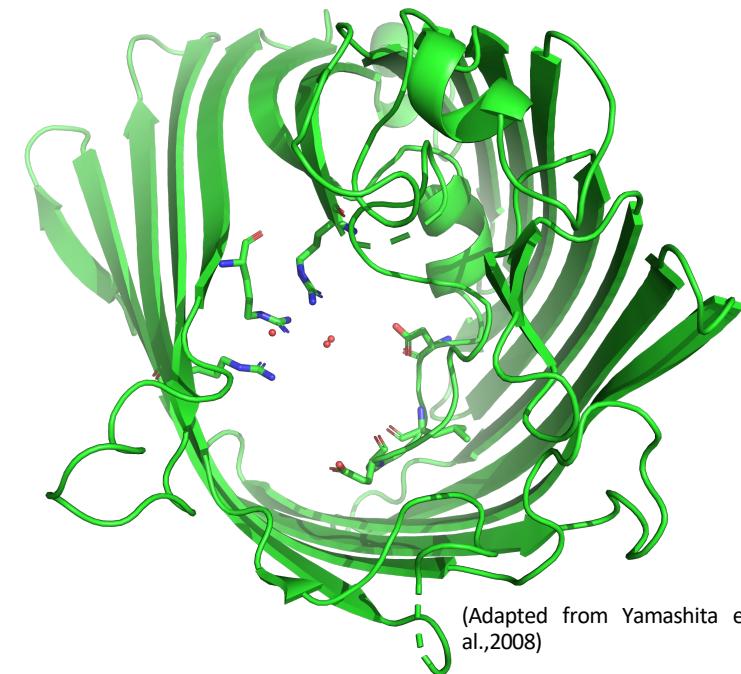
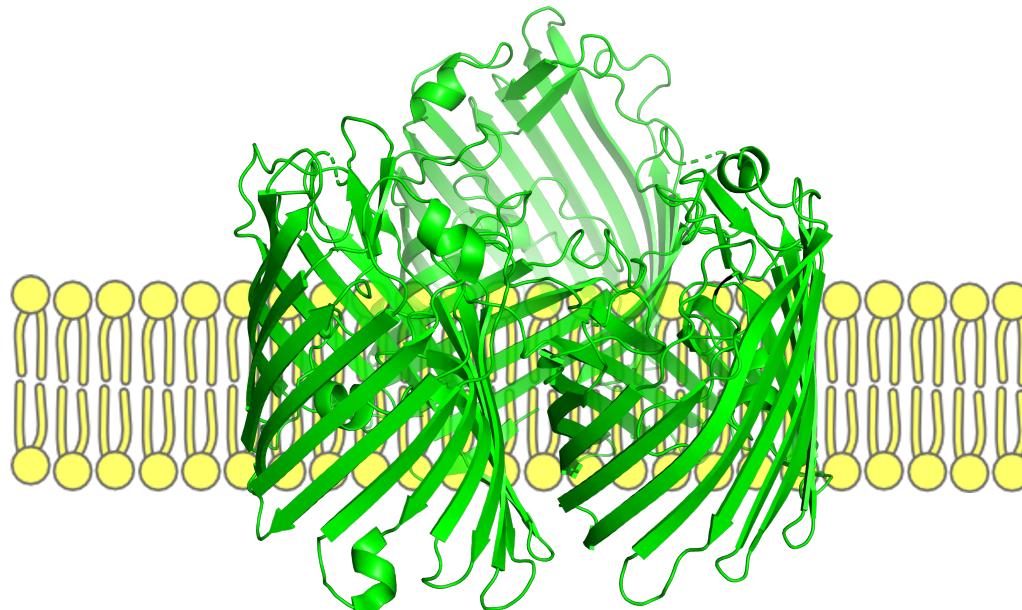
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**Labelling can be done partially by H/D exchange or fully by producing deuterated protein**

# OmpF: Model membrane protein system



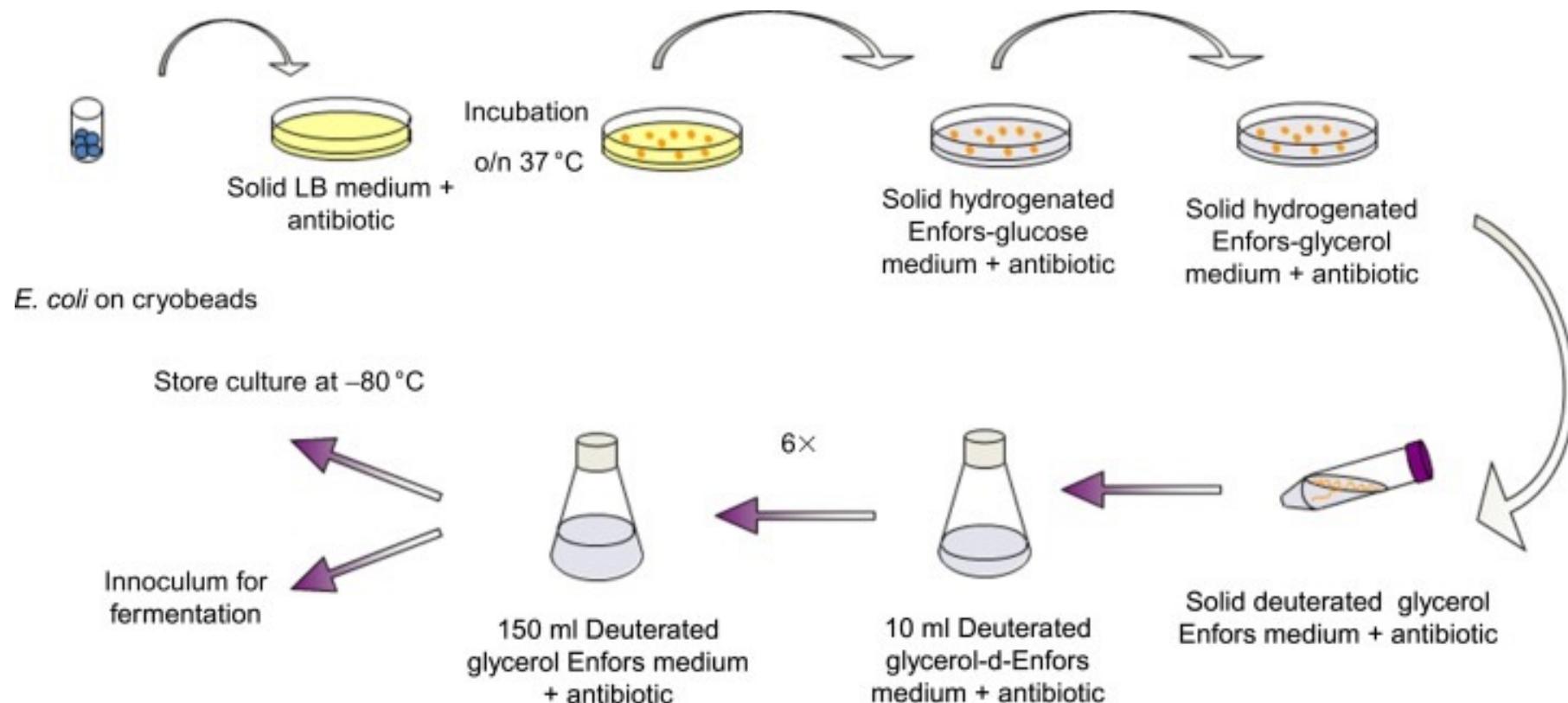
## Characteristics

- It constitutes around 7% of the total protein mass of the bacteria
- X-Ray structure at 1.8 Å° resolution
- Well studied and highly stable membrane protein

## Research Interest

- Hydrogen bonding between each monomer in the trimeric structure
- Understand the flexibility of water channel that involves a network of acidic and basic residues
- Potential role in drug resistance of bacteria

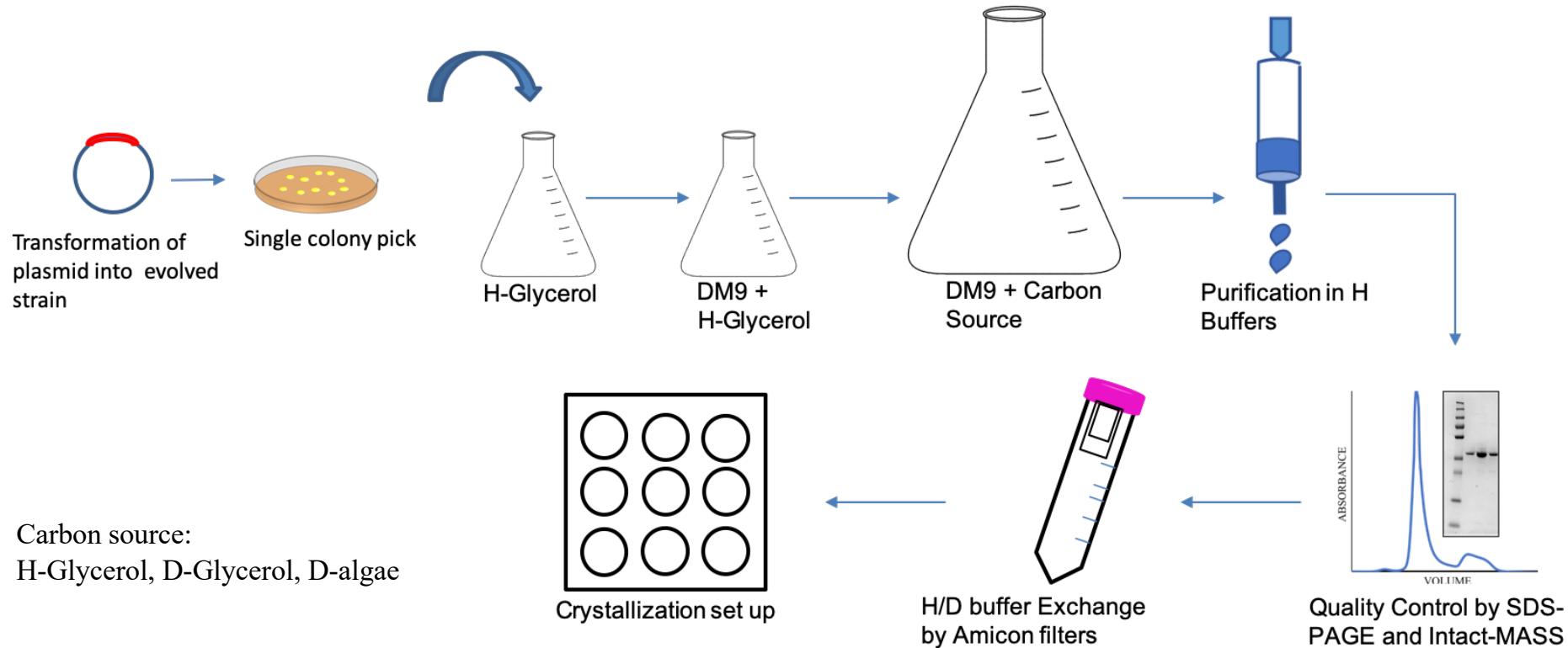
# Current adaptation Strategy to produce deuterated proteins



Adapted from Haertlein et al, 2015

*E. coli* as the preferred host

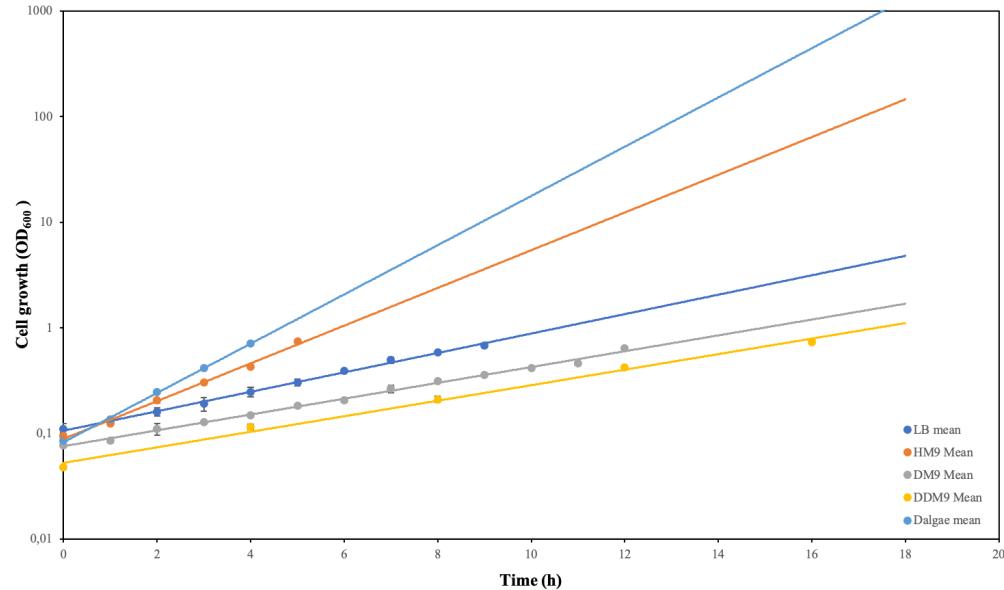
# Production of Deuterated proteins in *E. coli* using adapted strain



## Major requirement

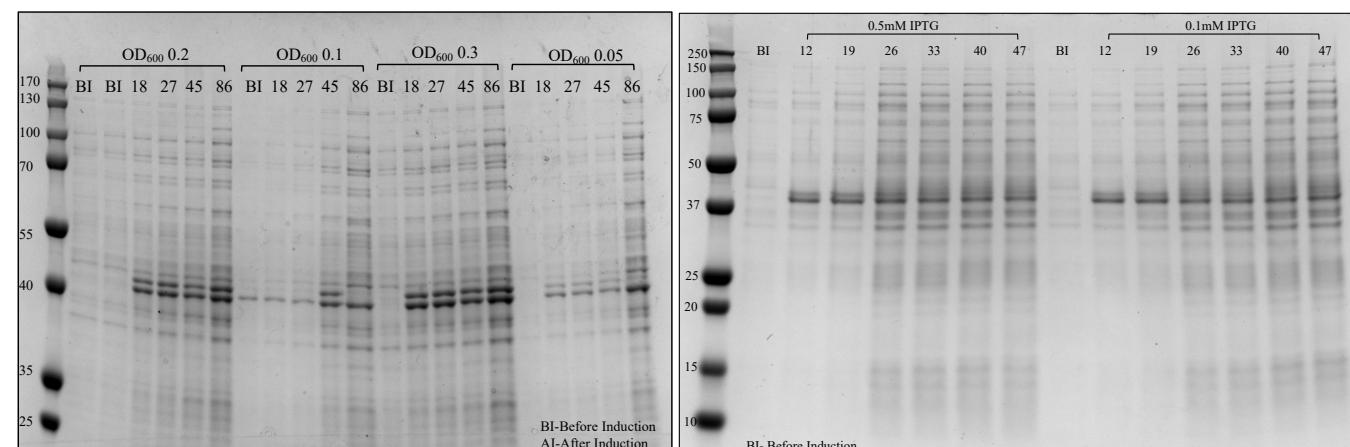
- Kanamycin resistance plasmid
- T7 RNA Polymerase

# Expression trials are necessary



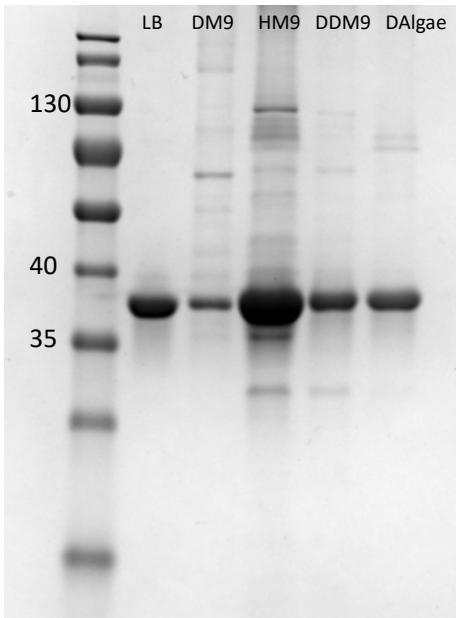
Growth Curve *E.coli* DA1(DE3) transformed with *ompF* in pET-24(+) grown in minimal media

- Bacterial growth in deuterated media is significantly reduced
- Optimal expression time is important
- Long incubation time had no effect on protein yield
- Starting culture optical density had a significant role in protein yield



*ompF* expression level test in deuterated minimal media

# Does labelling has significant effect on membrane protein stability?



SDS-PAGE analysis of purified OmpF protein samples produced using different media components

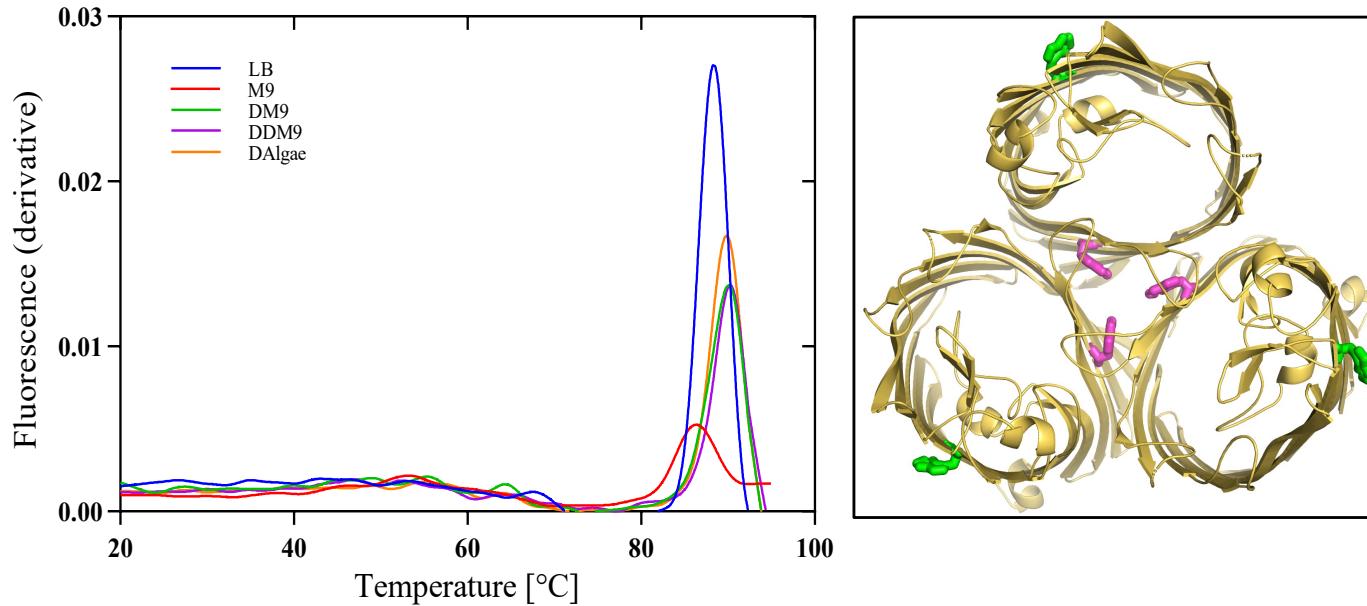
OmpF yield comparison in different media

Growth media used	Protein yield (mg/l)
LB	1.5
M9	1
DM9	1
DDM9	1
DAlgae	1

Theoretically and experimentally calculated mass of hydrogenated and deuterated OmpF in different media components.

Deuterated Protein Sample	Calculated Molecular weight (Da)	Mass from MS (Da)	% Deuterated
DM9 (H-Glycerol)	39539.6	38759.7	90
DDM9 (D-Glycerol)	39539.6	39054.8	100
Dalgae	39539.6	38860.7	99

# Does labelling has significant effect on membrane protein stability?

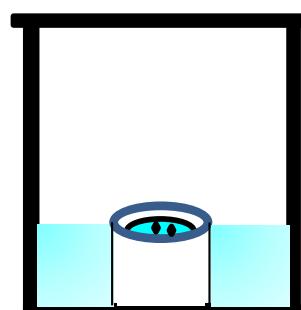
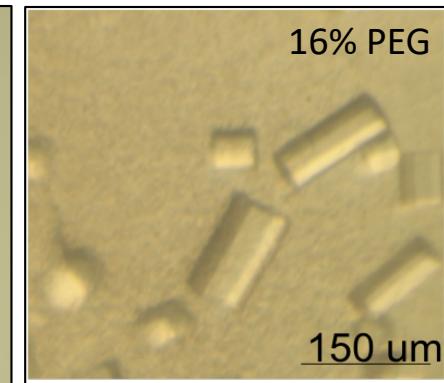
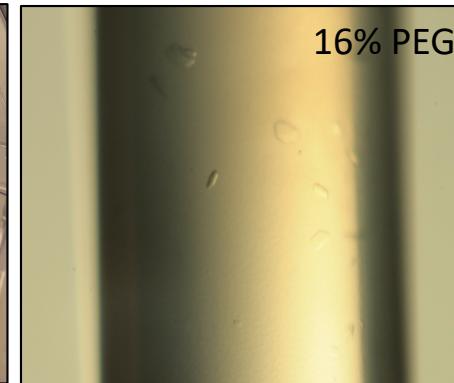
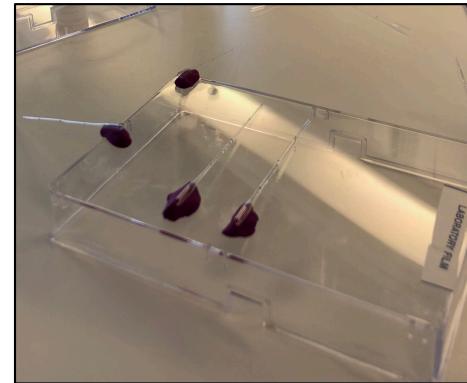
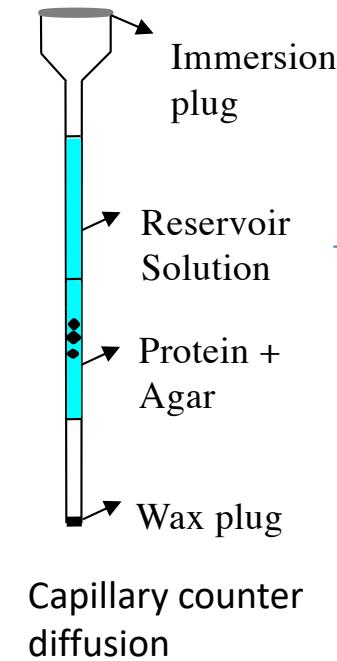


- Thermal unfolding temperature was higher for deuterated protein samples
- Temperature determines oligomeric structure of protein

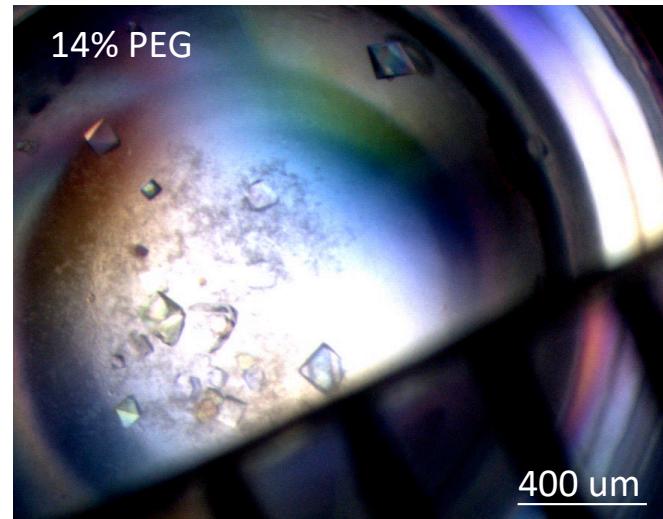
Protein Sample	Protein Unfolding Temperature (°C)
LB	88.3
M9	86.5
DM9	90.0
DDM9	90.2
DAlgae	89.9

Protein unfolding temperatures ( $T_m$  in °C) as determined by Nano-DSF

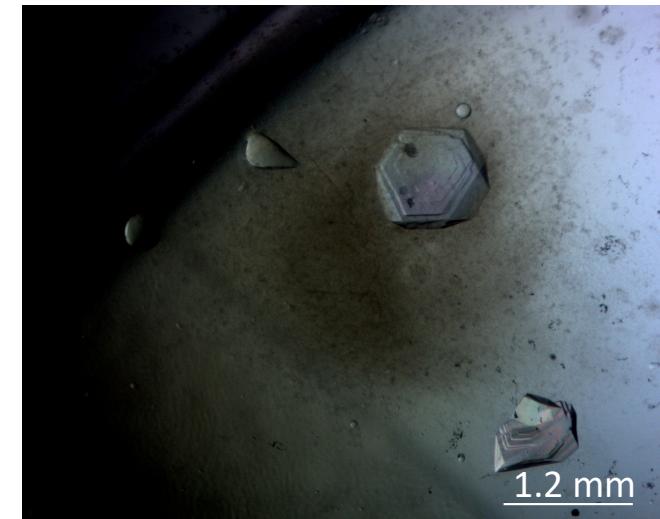
# Techniques used for growing large crystals



Vapour diffusion



H/D Exchange

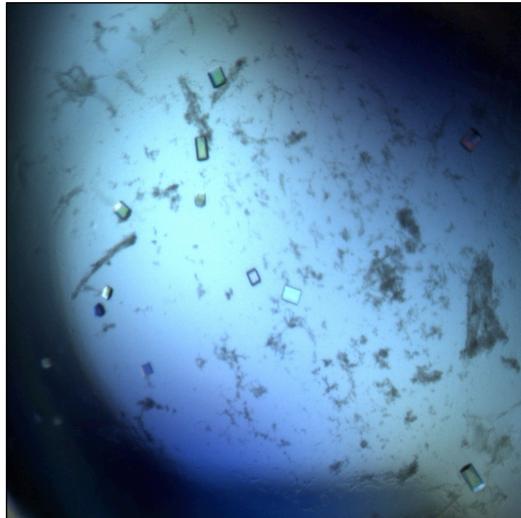


Hydrogenated

# Crystallization trials of OmpF in heavy water using vapor diffusion



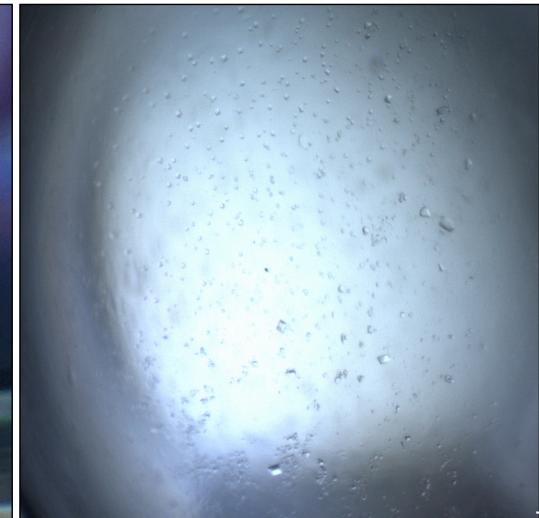
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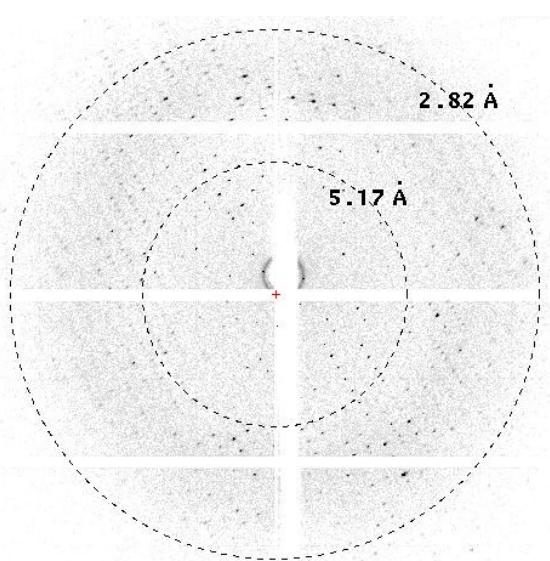
HM9



DM9



DDM9



?

# Conclusions

- The protocol developed in this study serves as a convenient way to produce and improve the yield of perdeuterated membrane protein without using large fermenters.
- Perdeuteration has subtle effects upon the physicochemical properties of proteins.
- Deuteration affects the secondary structure motifs and hydrophobic surface exposure during the unfolding events.
- Further challenges to reoptimise crystallization conditions in D<sub>2</sub>O.
- The limitations of large crystal size and small unit cell volumes might not hold with the construction of ESS in Lund, Sweden.

# Acknowledgements



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RAMP Members



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Thank You for your attention!!! 😊



Questions and  
Suggestions??